

Chemical Modification of Radiation Response

Oxygen

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Oxygen – best known and most general radiosensitizer

- The slopes of survival curves for cells exposed to sparsely ionizing radiation in hypoxia and in well oxygenated environments differ by about a factor of 3 – the oxygen effect.
- Hypoxia means low oxygen, anoxia means no oxygen.

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Oxygen Enhancement Ratio (OER)

- $D_0(\text{hypoxia})/D_0(\text{oxygenated})$
 - = dose(hypoxia)/dose(oxygenated) for the same effect
- If the survival curves in both air and hypoxia extrapolate back to the same n value, the curves are said to be purely dose modifying.
- OER is usually about 3 at high radiation doses, but often has a lower value of about 2 at low doses (at or below 2 Gy).

How much O₂ is required?

Survival curves show that relatively little O₂ is needed, e.g., as little as 100 ppm O₂ (0.075 mm Hg) causes significant sensitization compared to the response in anoxia.

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“K-curve” – plot of relative radiosensitivity vs. oxygen concentration – shows half-maximal effect of oxygen at about 0.5% (3 mm Hg) O₂. (For comparison, venous p O₂ is about 50 mm Hg and arterial is about 100 mm Hg; air is 155 Hg.)

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Timescale – When must O₂ be present?

- Very fast response techniques show that O₂ must be present during irradiation or added within milliseconds after irradiation in order to be sensitizing.
- For most practical purposes, it must be present during the radiation exposure.

Mechanism

- Mechanism(s) of the oxygen effect really not know, although, clearly, O₂ acts at the free radical level.
- The reactions involved may be:
$$\text{O}_2 + e^-_{\text{aq}} \rightarrow \text{O}_2^{\cdot-} \quad \text{or} \quad \text{O}_2 + \text{R}^\cdot \rightarrow \text{R O}_2^\cdot$$
- The later reaction is sometimes called “fixation” of damage in the lethal form and occurs in competition with chemical repair of damage, perhaps by H atom donation from thiols (to be discussed more below).

Importance of the oxygen effect

- Thomlinson and Gray (1955) studied sections of bronchial carcinoma
- Small tumors (<160 μ) – no necrosis
- Tumors over 200 μ - necrotic centers surrounded by sheath of healthy cells
- Sheath of growing cells always 100-180 μ
- They also calculated O₂ diffusion in tissues and found that all O₂ should be metabolized at a distance of 150-200 μ from a capillary, in good agreement with the observations of necrosis.
- Actually, there will be an O₂ concentration gradient through the tumor, so some tumor cells will have enough O₂ to grow but will be radiobiologically hypoxic (and therefore radioresistant). These cells may limit the effectiveness of radiation therapy of tumors.

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This model is really a gross oversimplification of tumor oxygenation, but emphasizes the importance of oxygen in radiation therapy and explains why a great deal of research has been conducted into ways to overcome hypoxic cells.

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Hypoxic cells may be of two types:

Diffusion-limited – as described by Thomlinson and Gray (chronic hypoxia)

Perfusion-limited – cells intermittently hypoxic only when the blood flow transiently stops on their vessel (acute hypoxia).

Dealing with the different types of hypoxia may require different methods.

Do hypoxic cells really exist in tumors?

The first demonstration that they do exist in an experimental animal tumor was made by Powers and Tolmach using the dilution assay technique. They observed a two component survival curve:

Low doses – $D_0 = 1.1$ Gy – normal

high doses - $D_0 = 2.6$ Gy

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The ratio of about 2.5 between two D_0 values suggested the OER: the low dose component was from oxygenated cells and the high dose component from hypoxic cells.

Back-extrapolation of the high dose component to the y-axis gives the % hypoxic cells in the tumor.

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Hyperbaric Oxygen (HBO)

Shortly after the identification of hypoxia as a potential cause of tumor radioresistance, clinical trials were begun with hyperbaric oxygen.

Most trials have been relatively small and used unconventional fractionation patterns, but in several there was to be an advantage, albeit small, to the use of HBO.

Problems included:

- Questions of whether increases in dissolved O₂ in plasma really resulted in increases in hypoxic tumor cells.
- Some normal tissues may be of sufficiently low O₂ to be sensitized by the HBO.
- Practical problems such as patient convulsions due to oxygen, patient complications in lungs and ears, claustrophobia, danger of fire and explosion, etc.

Use of carbogen (95% O₂/5% C O₂ at 1 atm) with or without perfluorochemicals may give as good or better results than HBO.

Evidence for the presence of hypoxic cells in human tumors:

Tumor histology

Oxygen electrode measurements

Clinical gains with hyperbaric oxygen

Studies showing anemia is poor prognostic factor also associated with local failure, but pre-transfusion help.

Radiation Sensitizers

Radiosensitizers

- Agents which enhance the response of cells to radiation.

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- Ideally, radiosensitizers would selectively sensitize tumor cells while having no effect on normal tissues.

Non-hypoxic cell radiosensitizers

- Halogenated pyrimidines, BUdR and IUdR
- Are incorporated into DNA in place of thymine. Therefore, the tumor cells must be cycling faster than the nearby dose-limiting normal tissues.
- IUdR and BUdR have similar sensitization with X-rays, but IUdR is preferable clinically because it sensitizes cells much less to fluorescent light, so less harmful side effects.
- Sensitize both hypoxic and oxygenated cells.
- The degree of sensitization depends on the amount of halogenated pyrimidine incorporated into a cell.
- Clinical trials with IUdR are underway with gliomas and sarcomas with encouraging early results. The agent is being delivered with brachytherapy as well as external beam therapy.

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Hypoxic cell radiosensitizers

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Hypoxic cell radiosensitizers – electron-affinic compounds that selectively sensitize hypoxic cells while having no effect on oxygenated cells.

Ideal properties of sensitizer:

- Selectively sensitize hypoxic cells
- Chemically stable and slowly metabolized
- Highly soluble in water and lipids so can diffuse to hypoxic tumor cells
- Effective throughout cell cycle
- Effective at low daily doses of radiation

$SER = D_0(\text{without sensitizer})/D_0(\text{with sensitizer})$

Nitroimidazole class of compounds most studied.

- Metronidazole (flagyl – a 5-nitroimidazole) gave *in vitro* SER = 1.7 and *in vivo* ER = 1.3.

- Misonidazole (Ro-07-0582 – 2 – nitroimidazole) better sensitizer, with *in vitro* SER = 2.5 for hypoxic cells (no effect on oxygenated cells) and tumor SER up to 1.8.

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Hypoxic cell cytotoxins; Bioreductive agents (quinines, nitro compounds, benzotriazine di-N-oxides)

- Drugs that are preferentially toxic to hypoxic cells.

<u>Agent</u>	<u>Hypoxic Cell Cytotoxicity Ratio*</u>
Mitomycin C	2
EO9	5
Metronidazole	2
Misonidazole	11
Nitrofurazone	8.5
RSU 1069	67
SR4233 (tirapazamine)	50

* Drug dose required to kill given proportion of aerobic cells divided by that needed to kill same fraction of hypoxic cells

Data all for V79 cells; HCR values vary with cell line

- Killing hypoxic cells may have greater therapeutic advantage than radiosensitizing them because:
- hypoxic cytotoxins kill cells resistant to radiation and most chemotherapy, producing complementary cytotoxicity.
- random fluctuations in acute hypoxia could create a situation where hypoxia could be used to advantage.

Modeling studies show that if a hypoxic cytotoxin is given with every dose fraction, the overall kill in a hypoxic tumor can be greater than if the tumor is fully oxygenated. This occurs when the drug kills at least 50% of the hypoxic cells each time it is given.

However, for a hypoxic cytotoxin to be effective in a fractionated regimen, there must be rehypoxification between fractions.

Results of clinical trials have been disappointing.

Table 1. Results of randomized controlled trials of radiosensitizing methods

	<u>Hyperbaric Oxygen</u>	<u>Metronidazole</u>	<u>Misonidazole</u>
Therapeutic Benefit	3	0	2
Significantly Improved Results	0	2	4
Margin in favor (not significant)	6	0	2
No Difference	6	4	30
Margin Against (not significant)	0	0	1
Adverse Response	0	0	0
	15	6	39

(from Dische in Malaise *et al* 1989)

Possible reasons for failure of misonidazole in clinical trials include:

- Relatively small sample size and heterogeneous population may have precluded observation of a small effect.
- SER may be lower at clinically relevant doses.
- Misonidazole has cumulative neuropathy, which limited the dose that could be given and the number of fractions with which it could be given. (Total of 12 g/m², so with single dose of 2 g/m², which might be expected to give ER in hypoxic cells of about 1.3-1.4, could only give misonidazole with 6 fractions.)

Radioprotectors

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Radioprotectors – agents which decrease the response of cells to radiation.

Best radioprotectors are thiols

Dose Reduction Factor (DFR) = Protection Factor (PF) =

$$\frac{\text{dose of radiation in the presence of the drug}}{\text{dose of radiation in the absence of the drug}}$$

to produce a given level of effect.

Proposed mechanisms for radioprotection by thiols:

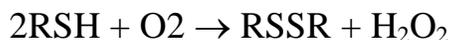


The rate of the scavenging reaction is independent of the presence or absence of oxygen, so scavenging will not explain the differential protection by thiols in hypoxia and air.

Donation of H atoms to organic radicals, in competition with damage “fixation” of those radicals by oxygen



Consumption of oxygen, so hypoxia is produced



WR2721 (amifostime) and related compounds

Covering the SH with a phosphate group decreased the toxicity

TABLE 9.1. *Effect of adding a Phosphate-Covering Function on the Free Sulfhydryl of β -Mercaptoethylamine (MEA)*

Drug	Formula	Mean 50% Lethal Dose (Range) in Mice	Dose-reduction Factor
MEA	$\text{NH}_2\text{-CH-CH}_2\text{-SH}$	343 (323-364)	1.6 at 200 mg/kg
MEA-PO ₃	$\text{NH}_2\text{-CH}_2\text{-CH-SH}_2\text{PO}_3$	777 (700-864)	2.1 at 500 mg/kg

In 1969 Yuhas and colleagues reported that WR2721 could protect normal tissues with less protection of tumors.

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Note that the degree of protection depends on the normal tissue type.

TABLE 9.2. Three Protectors in Practical Use

Compound	Structure	Use
WR-638	$\text{NH}_2\text{CH}_2\text{CH}_2\text{SPO}_3\text{HNa}$	Carried in field pack by Russian army (cystaphos)
WR-2721	$\text{NH}_2(\text{CH}_2)_3\text{NHCH}_2\text{CH}_2\text{SPO}_3\text{H}_2$	Protector in radiotherapy and carried by US astronauts on lunar trips (amifostine)
WR-1607	$\text{CH}_3(\text{CH}_2)_9\text{NHCH}_2\text{CH}_2\text{SSO}_3\text{H}$	Marketed as rat poison (d-CON)

Comparison of Hematopoietic and Gastrointestinal Dose Reduction Factors in Mice for the Three Compounds Listed Above

Compound	Drug Dose, mg/kg	Dose-reduction Factor	
		7 Days	30 Days
WR-638	500	1.6	2.1
WR-2721	900	1.8	2.7
WR-1607	10	—	2.1

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However, radioprotection by WR2721 is not restricted to normal tissues; tumors can be protected.

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Some factors which affect the degree of radioprotection of normal tissue or tumor by WR2721 include:

- Rate and extent of uptake of WR2721, which depends on:
- Concentration of alkaline phosphatase in plasma membrane pH
- Oxygen concentration
- Endogenous thiol level
- Fractionation pattern (single versus multiple doses)

Clinical trials have shown side effects such as hypotension, nausea, vomiting, and hypocalcemia. Hypotension has been the dose limiting toxicity.

- Only a few clinical trials with radiation have been undertaken. Although acute reactions to radiation appear to be protected, the effects on late radiation reactions remain to be evaluated in long term studies. Acute reactions do not always predict late reactions, so dose escalation must proceed cautiously.
- Clinical trials suggest amifostine may be useful to protect against nephro-neuro- and oto-toxicity from *cis*-platin treatment and cyclophosphamide-induced granulocytopenia.
- More recently, interest in WR2721 and its derivatives has centered on observations that these drugs effectively protect against radiation-induced and some drug-induced mutations and neoplastic transformation.

<u>Protector</u>	<u>Treatment</u>	<u>Endpoint</u>	<u>PF</u>
WR-1065	Gamma rays	HGPRT mutations	5.1
WR-1065	Neutrons	HGPRT mutations	3.3
WR-1065	<i>cis</i> -PT	HGPRT mutations	7.1
WR-1065	HN ₂	HGPRT mutations	3.4
WR-1065	BLM	HGPRT mutations	2.8
WR-1065	gamma rays	Transformation	6.0
WR-2721	gamma rays	Preneoplastic lesions	9.7
WR-2721	gamma rays	Tumor induction	3.1
Mixture	X-rays	Tumor induction	1.4

From Grdina *et al*

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