

Lecture 35
5/12/04

Review Sunday 1-4pm room 2-147
TD review starts at 2:30

Continued from lecture 34 (see cartoon diagram)
Degradation through polyubiquitination- (eukaryotes)

Specificity comes from E3 (remember ~100 E3's inside the cell)
You could imagine one type of E3 would interact with positively charged N-terminal amino acid, and another would recognize hydrophobic N-term.
Several ubiquitins are added on to a lysine on the protein by E3 and E2
Ubiquitination may have many roles inside the cell- we are only discussing its role in protein degradation in yeast.

Roles of E1,E2,E3 in ubiquitination (See handout 4d for cartoons of all of this)

- 1) ATP activated Ub through adenylation
- 2) E1 binds covalently to C-term of Ub through a thioester
Need fewer E1-> higher turnover rate than E2, E3
- 3) Ub transferr to E2, again as a thioester
- 4) Substrate needs N-end amino acid + lysine-> E3 interacts with substrate and E2 to catalyze attachment of Ub to lysine through isopeptide linkage
- 5) Next Ub is added onto the lysine 48 of the previous Ub (E2, E3 repeat same process until 3-4 Ub are added)
- 6) PolyUb is recognized by the 19S cap to human proteosome and the substrate is degraded
- 7) Ub can be cleaved off intact and then reused

Example of proteins targeted for degradation (regulation through rapid degradation)
TF= transcription factor- heterodimer of alpha and beta
Acts as “oxygen sensor”, when there is a lack of oxygen, TF activates transcription of a # of genes
Can stimulate angiogenesis- new blood vessel growth to find more O₂

To deactivate this transcription, degrade 1 of the 2 subunits

In the presence of normal O₂, there is post-translational modification to hydroxylate a proline on the alpha subunit. This interacts with E3, targets alpha for degradation by the proteosome

This is how the system is capable of “oxygen sensing” without O₂, the reaction can’t occur, alpha is not degraded, transcription is activated

Quick 5.08 Summary

Biochemistry

-amazing complexity

-must relate in vitro studies to in vivo reality

-Macromolecular machines- conserved through 3 kingdoms (use simpler bacterial system as a starting point for understanding)

-FAS, PKS, NRPS

-Ribosome, translation

-“Foldosomes,” “Proteosomes”

Methods

- 1) reconstitution – self-assembly
- 2) chemical probes (photoaffinity labeling, crosslinking, fluorescence)
- 3) site directed mutagenesis, unnatural amino acids
- 4) natural product inhibitors- freeze conformation
- 5) cryoEM, x-ray, NMY structure
- 6) kinetics (dynamics) pre-steady state and steady state
- 7) electrophoresis
- 8) use of radioactivity
- 9) mass spectrometry
- 10) FRET