Recitation 4-9-14

EF Lectures #14 & 15
Protein Interactions & Gene Networks

Announcements

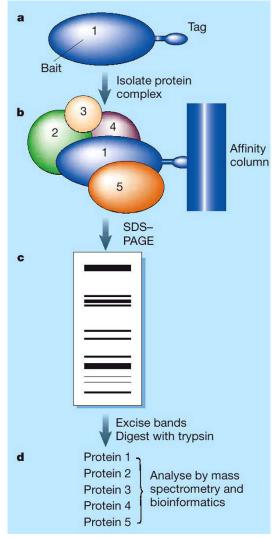
- Problem Set 4 due next Thursday (April 17)
- Project write-up due Tuesday, April 22

Outline

- Experimenta methods to detect protein interactions
 - Affinity Purification
 - Tandem Affinity Purification (TAP)
 - Mass Spectrometry
 - Yeast two-hybrid
- Bayesian Networks
- Clustering methods
 - Hierarchical clustering
 - K-means clustering
- Linea Regressio Mutual information

Affinity Purification

- To detect interaction partners of a protein of interest (bait), the bait is tagged by introducing protein-tag DNA construct into cells. Once the construct is expressed and incorporated into cellular complexes, the tag is used to pull down other interacting proteins, by Mass Spectrometry.
- Can do this for every protein to analyze proteins on a proteome-wide scale.
- Fairly high (~30% for 2002 yeast genome-wide study) False Positive Rate with single-affinity purification, but also some False N known interactors & comple



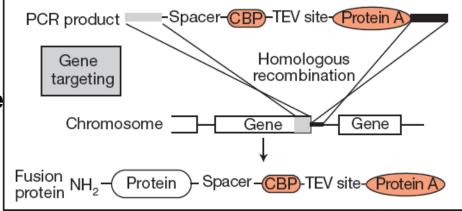
Courtesy of Macmillan Publishers Limited. Used with permission. Source: Kumar, Anuj, and Michael Snyder. "Proteomics: Protein Complexes take the Bait." *Nature* 415, no. 6868 (2002): 123-4.

Tandem Affinity Purification (TAP)

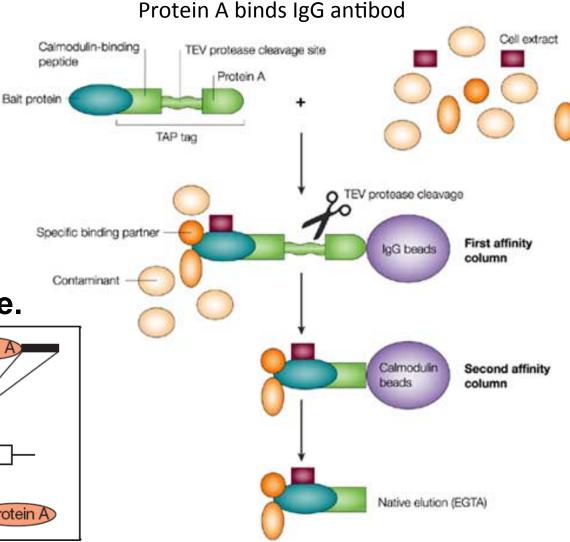
-To cut down on false positives, two affinity purification steps

 However, fewer false positives likely means more false negatives

Gavin et al. (2002) Nature.



Courtesy of Macmillan Publishers Limited. Used with permission. Source: Gavin, Anne-Claude, Markus Bösche, et al. "Functional Organization of the Yeast Proteome by Systematic Analysis of Protein Complexes." *Nature* 415, no. 6868 (2002): 141-7.

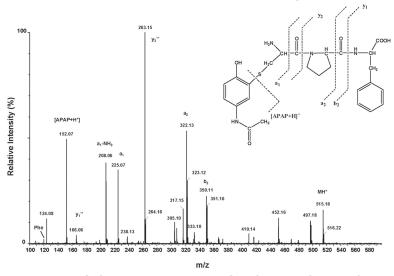


Nature Reviews | Molecular Cell Biology

Courtesy of Macmillan Publishers Limited. Used with permission. Source: Huber, Lukas A. "Is Proteomics Heading in the Wrong Direction?" *Nature Reviews Molecular Cell Biology* 4, no. 1 (2003): 74-80.

Mass Spectrometry (MS)

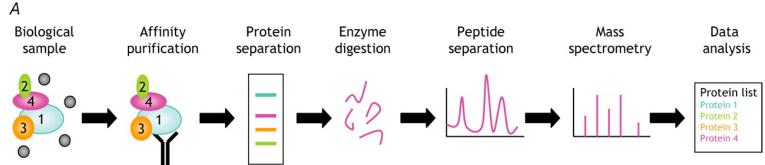
- Analytical technique that produces spectra of the masses of atoms or molecules that comprise a sample
- Works by ionizing chemical compounds to generate charged molecules & measuring the mass-to-charge (m/z) ratio



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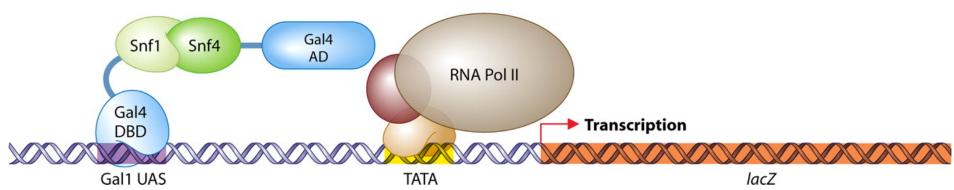
Source: Damsten, Micaela C., Jan NM Commandeur, et al. "Liquid Chromatography / Tandem Mass Spectrometry Detection of Covalent Binding of Acetaminophen to Human Serum Albumin." *Drug Metabolism and Disposition* 35, no. 8 (2007): 1408-17.



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Yeast Two-Hybrid (Y2H)

- Used to detect interactors with your bait protein of interest
- Introduce two plasmids into yeast cells:
 - 1. DNA-binding domain (DBD) Bait fusion (in this case, Snf1 is bait)
 - 2. Prey Activator domain (AD) fusion (in this case **Snf4** is prey)
 - AD needed to recruit PolII for transcription of reporter gene
 - Often will screen a library of potential prey molecules
 - This example uses the well characterized Gal4 transcription activator protein in yeast.
 - lacZ transcription can be detected by colorimetric inspection (can use other reporter genes such at metabolic enzyme (His production) and growth on minimal media lacking His)
- Y2H will miss interactions for prey proteins that are not soluble and/or don't localize to the nucleus
- But can detect more transient interactions that may not be captured by affinity purification



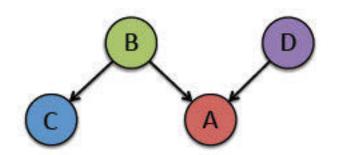
© American Society for Microbiology. All rights reserved. This content is excluded from our Creative Commons license. For more information, see http://ocw.mit.edu/help/faq-fair-use/. Source: Stynen, Bram, Hélène Tournu, et al. "Diversity in Genetic in Vivo Methods for Protein-Protein Interaction Studies: From the Yeast Two-hybrid System to the Mammalian Split-luciferase System." *Microbiology and Molecular Biology Reviews* 76, no. 2 (2012): 331-82.

- If we have 3 binary variables A, B, C that we can observe, how many variables do we need to fully specify joint probabilit P(A=a,B=b,C=c) in the following situations:
 - A,B,C are all independent of each other?
 - $P(A=a,B=b,C=c) = P_A(a) P_B(b) P_C(c)$ 3 parameters (more generally, n for n binary variables since 1 probability (prob. of ON) needed for each)
 - Cannot assume any independencies?
 - Need all possible combinations of A,B,C = 2^3 -1 (= 2^n -1 for n binary variables since there are 2^n combinations, but last one is determined since all probabilities must sum to 1)
 - The Bayesian network tells us about independencies between variables, and allows us to factor the joint probability accordingly
- A Bayesian networks is a way of representing a set of random variables and their conditional dependencies. Consists of:
 - 1. Directed (acyclic) graph over the variables
 - 2. Associated probability distributions:
 - Prior probabilities of all root nodes and
 - Conditional probabilities of all child nodes given their parents

- The directed graph consists of:
 - Nodes = random variables (events)
 - Edges indicate dependencies between variables
- Then the distribution of a random variable *depends only on its*Parent and child nodes: if there

parent nodes:

A Bayesian network with 4 nodes



Parent and child nodes: if there is directed edge starting from *i* and ending at *j*, then *i* is a parent of *j* and *j* is a child of *i*

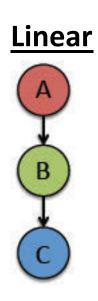
- C has parent B, B has child C
- A has parents B and D

Root nodes = nodes with no parents (no incoming edges) – here B and D Leaf nodes = nodes with no children – here A and C

Need the following probabilities to fully specify the model:

- Prior probabilities of all root nodes = P(B) and P(D)
- Conditional prob. of all child nodes given parents = P(C|B) and P(A|B,D)

There are 3 types of connections that can occur between a random variable B and its immediate neighbors A and C:



Factor P(A,B,C) according to the independencies indicated in this graph:

P(A,B,C) = P(A)P(B|A)P(C|B)

Are A and C independent if B is <u>unknown</u>?

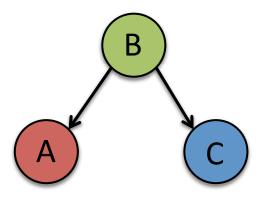
No – if B is unknown, then knowing A tells us something about C (through unknown B)

Are A and C independent if B is known?

Yes – if B is known, there is no further information in A about C

There are 3 types of connections that can occur between a random variable B and its immediate neighbors A and C:

Diverging



Example of this:

B is the bias of a coin, and **A** and **C** are the outcomes of independent flips of that coin

Factor P(A,B,C) according to the independencies indicated in this graph:

P(A,B,C) = P(B)P(A|B)P(C|B)

Are A and C independent if B is <u>unknown</u>?

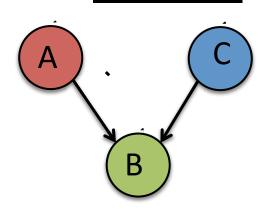
No – if B is unknown, then knowing A tells us something about C (through unknown B)

Are A and C independent if B is known?

Yes – if B is known, there is no further information that A can tell us about C

There are 3 types of connections that can occur between a random variable B and its immediate neighbors A and C:

Converging



Example of this:

A and C are two independent coin flips, B checks whether the resulting values are the same

Factor P(A,B,C) according to the independencies indicated in this graph:

P(A,B,C) = P(A)P(C)P(B|A,C)

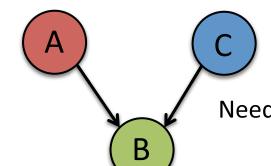
Are A and C independent if B is <u>unknown</u>?

Yes – if B is unknown, then knowing A tells us nothing about C

Are A and C independent if B is known?

No – if B is known, it tells us something about both A and C, so A and C are no longer independent

Converging



Given this graph structure, A and C are marginally independent (e.g. independent when B is marginalized out):

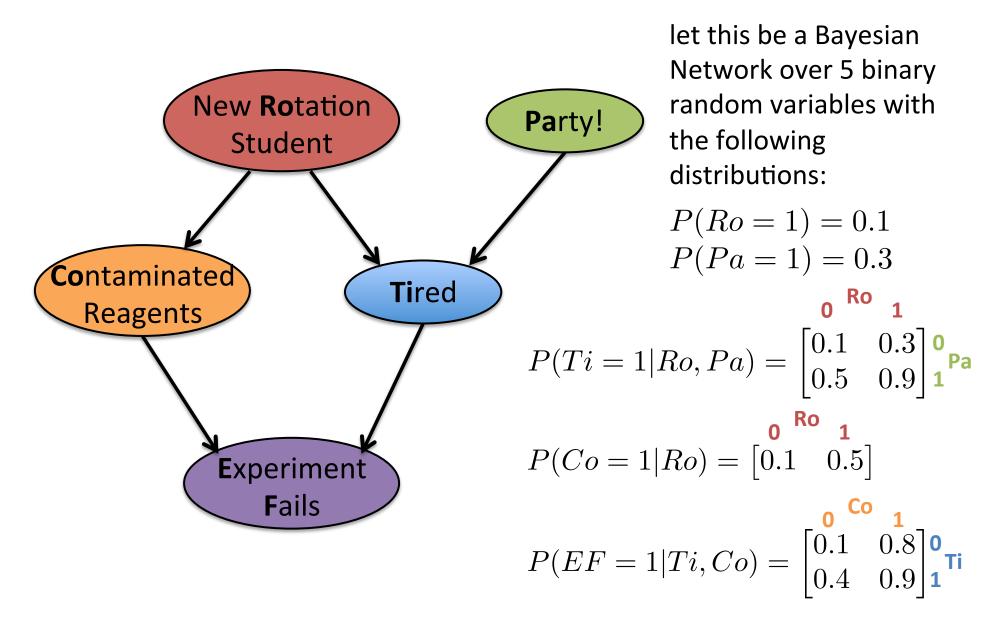
Need to show that
$$P(A,C)=P(A)P(C)$$

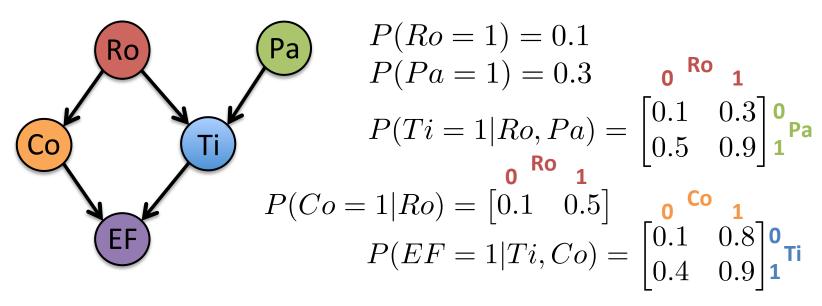
$$P(A,C)=\sum_B P(A,B,C) \text{ (marginalize out B)}$$

$$=\sum_B P(A)P(C)P(B|A,C)$$

$$=P(A)P(C)\sum_B P(B|A,C)$$

$$=P(A)P(C)$$

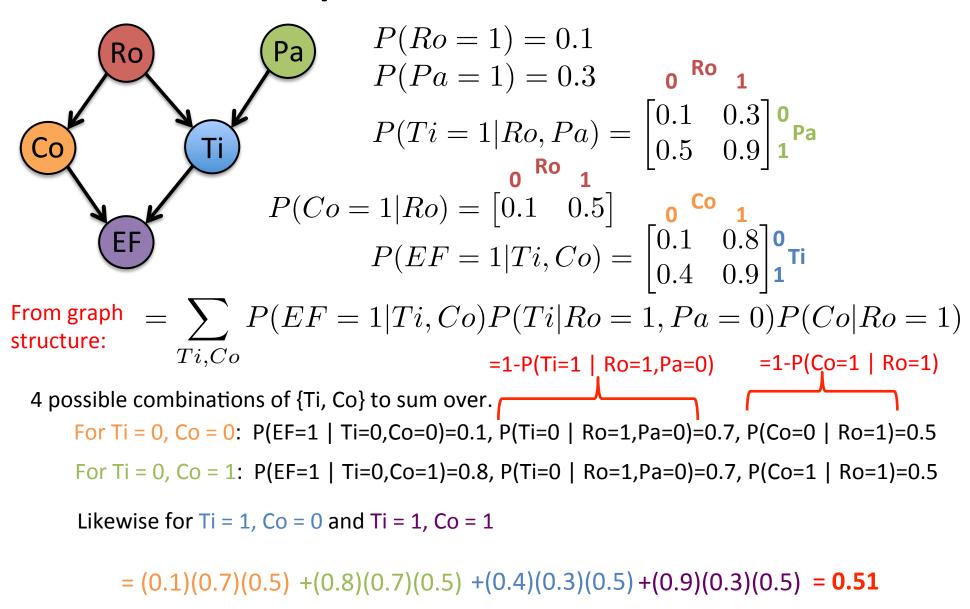




What is the probability that your experiment will fail given that there is a new rotation student, but there was no party last night? What is P(EF=1|Ro=1,Pa=0)?

$$P(EF = 1|Ro = 1, Pa = 0) = \sum_{Ti,Co} P(EF = 1, Ti, Co|Ro = 1, Pa = 0)$$

$$\frac{\text{From graph structure:}}{\text{structure:}} = \sum_{Ti,Co} P(EF=1|Ti,Co)P(Ti|Ro=1,Pa=0)P(Co|Ro=1)$$



Learning Bayesian Networks: parameters for given network

- Given a network structure (vertices and edges) and observations, we can learn the most likely conditional probabilities (e.g. we know a signaling pathway from previous experiments, but would like to determine its probabilities in response to a new stress condition)
 - This is an intference task, in contrast to the previous predictive task. Maximum Likelihood (ML) estimation – based on observed counts
 - find parameters (conditional probs.) that maximize the likelihood of the data:

$$\theta_{ML} = \underset{\alpha}{argmax} P(Data|\theta)$$

– example - given structure and observed counts below for binary vars A and B. estimate P(A) and P(B|A): n(A R)

(-)		2	· · ·		-8
$P(A=1) = (4+22)/(15+3+4+22) = 26/44 \approx 0.59$	(A)	0	0	15	
$P(B=1 A=0) = 3/(3+15) \approx 0.167$	Ţ	0	1	3	
$P(B=1 A=1) = 22/(22+4) \approx 0.846$	B	1	0	4	Ī
laximum <i>a posteriori</i> (MAP)	(b)	1	1	22	8

$$-\text{ incorporate prior knowledge }P(\theta) \text{about how params are distributed } \\ \hat{\theta}_{ML} = \underset{\theta}{argmax} P(\theta|data) = \underset{\theta}{argmax} \frac{P(data|\theta)P(\theta)}{P(Data)}$$

Observed counts plus pseudocounts corresponding to prior

Learning Bayesian Networks: network structure

 There are way too many possible structures for an exhaustive approach (e.g. trying every possible structure and calculating the likelihood of the data given that structure)

Common greedy approach (what Pebl does in Pset 4):

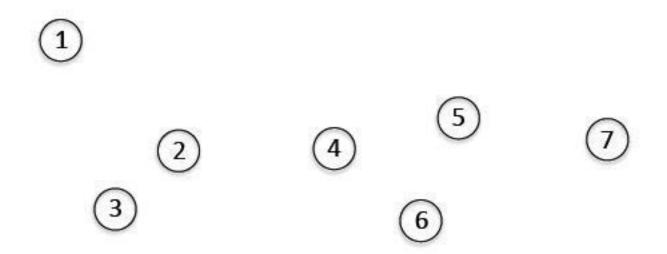
- start with a random network
- make a small perturbation (e.g. adding or removing an edge) and rescore network
- if network scores higher, accept (otherwise reject change)
- repeat from many starting points, pick best one

Simulated Annealing approach:

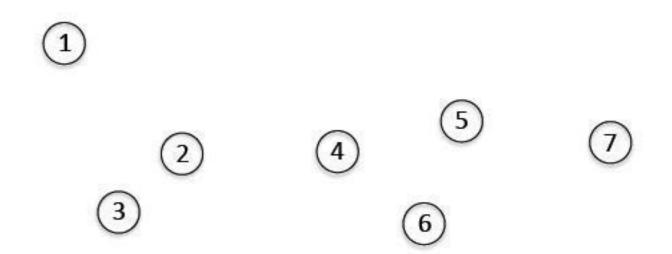
- similar to above, but accept lower scoring network with some probability proportional to difference in scores and temperature
- accept with higher probability initially, then "lower" temp gradually

- Useful when trying to find structure (e.g. clusters of genes upregulated in repsonse to a stress) in your data
- Algorithm:
 - initialize every point to be its own cluster
 - until only 1 cluster left:
 - calculate distance between each cluster and all other clusters : $O(N^2)$ for each connection -> $O(N^3)$ overall
 - find the two closest clusters, merge them into one cluster
- Can use various distance/similarity metrics (e.g. Euclidean distance, correlation, etc.)

 Let the following be 7 points in a 2-dimensional dataset – we want to do agglomerative hierarchical clustering on these points, using <u>Euclidean distance</u> as distance metric



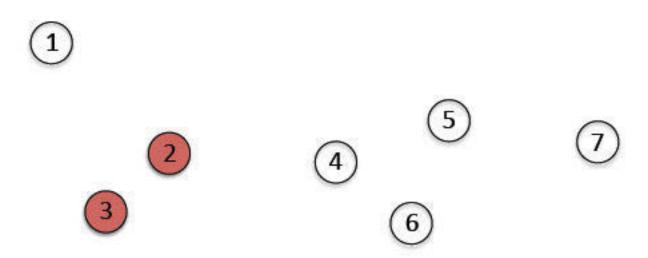
• (initialization) – initialize each point to be its own cluster



Build dendogram as we go to keep track of clusters – initially all nodes of dendogram are unconnected, connect them as we merge points into clusters



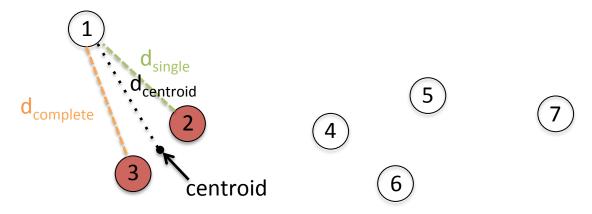
• (repeat until only 1 cluster left) – calculate distances between each pair of clusters, merge the two closest into single cluster



Closest are points 2 and 3 – merge these into a single cluster which we'll call Update dendogram:



- (repeat until only 1 cluster left) calculate distances between each pair of clusters, merge the two closest into single cluster
 - how do we do this for clusters with more than 1 point?



Let cluster **A** contain the set of points *i* and cluster **B** contains the set of points *j*, then the distance between **A** and **B** is:

Option (1): Single or complete linkage

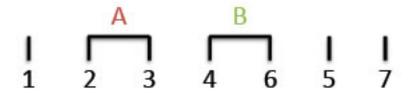
Calculate all distances d_{ij} between points i in A and all points j in other cluster B, and consider $dist(A,B) = min(d_{ij})$ for single linkage, $dist(A,B) = max(d_{ij})$ for complete linkage Option (2): **Centroid linkage**

For clusters A and B, compute the "centroid" or geometric center of the points in the cluster A_c and B_c , and dist(A,B) = dist(A_c , B_c)

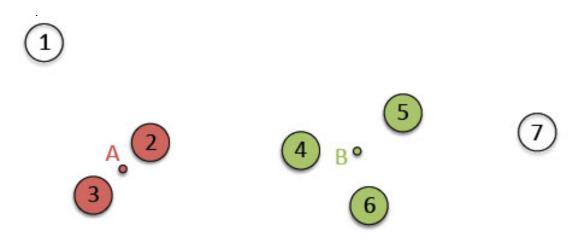
- (repeat until only 1 cluster left) calculate distances between each pair of clusters, merge the two closest into single cluster
 - use centroid linkage



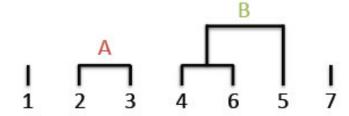
Closest clusters are points 4 and 6 – merge these into a single cluster B Update dendogram:



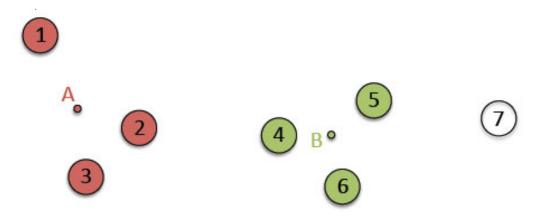
- (repeat until only 1 cluster left) calculate distances between each pair of clusters, merge the two closest into single cluster
 - use centroid linkage



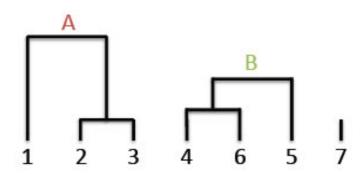
Closest clusters are B and 5 – merge these into a single cluster B Update dendogram:



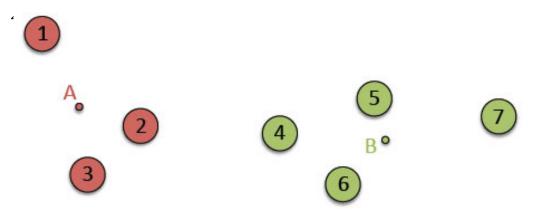
- (repeat until only 1 cluster left) calculate distances between each pair of clusters, merge the two closest into single cluster
 - use centroid linkage



Closest clusters
are A and 1 –
merge these into
a single clusterA
Update
dendogram:

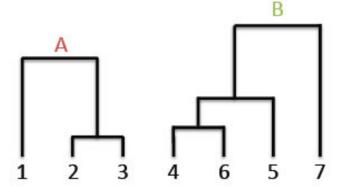


- (repeat until only 1 cluster left) calculate distances between each pair of clusters, merge the two closest into single cluster
 - use centroid linkage

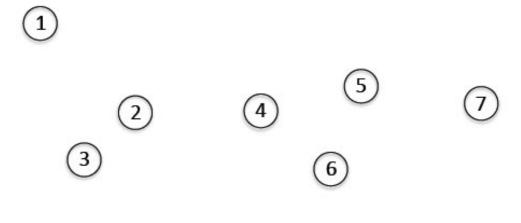


Closest clusters are
B and 7 – merge
these into a single
cluster B

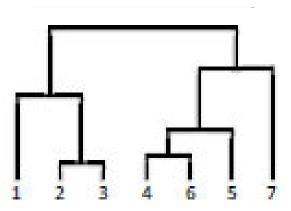
Update dendogram:



- (repeat until only 1 cluster left) calculate distances between each pair of clusters, merge the two closest into single cluster
 - use centroid linkage



Only two clusters left, merge them. Update dendogram:



Only one cluster remaining, so we're done!

- Can always cluster data, get a dendrogram and discover some "structure" in your data, but interpreting or assigning meaning to clusters is much more difficult
 - clusters may not corresponding to anything biologically meaningful
- In contrast to agglomerative ("bottom-up") clustering shown thus far, there is also divisive hierarchical clustering (topdown):
 - start with everything in one cluster, then cut the cluster into 2, then
 cut those clusters, etc., until you have the desired number of clusters

K-means clustering

- Goal: Find a set of k clusters that minimizes the distances of each point in the cluster to the cluster's mean
- You must a priori select k, the number of clusters to return
- Algorithm:
 - For all points X_i:
 - Assign X_i to the cluster with the closest mean
 - Recalculate the mean of each cluster based on previous iteration's assignments

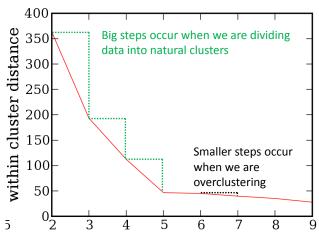
Repeat until convergence (no assignments change)

K-means clustering example (k=4)



K-means clustering

- Deterministic given:
 - 1. Choice of *k*
 - 2. The k starting points for the clusters
- For 2: Generally want to run many times with different starting points to obtain most robust partition
- For 1:
 - Try many different ks (below and above what you think it might be)
 - Intuitively, you should see large decreases in the intra-cluster distance when uncovering true underlying clusters & smaller decreases when overfitting



K-means clustering

- Deterministic given:
 - 1. Choice of k
 - 2. The k starting points for the clusters
- For 2: Generally want to run many times with different starting points to obtain most robust partition
- For 1:
 - Try many different ks (below and above what you think it might be)
 - Intuitively, you should see large decreases in the intra-cluster distance when uncovering true underlying clusters & smaller decreases when overfitting
 - Decision can be made automatically through frameworks such as Bayesian Information Criterion (BIC – penalizes addition of more free parameters; accepts model (i.e., k) that optimizes a tradeoff between increased likelihood of data from more clusters and increased number of free parameters)

Variations on K-means clustering

- Fuzzy k-means:
 - Rather than hard assignments (assigning each point to strictly 1 cluster), give soft assignments $u_{i,j}(\mu_{i,j})$ for all points $1 \le i \le N$, clusters $1 \le j \le K$
 - Constraint is $\sum_{i=1}^{n} \mu_{i,j} = 1$
 - Consider these soft assignments when recalculating the cluster means: $\hat{Y_j} = \frac{\sum_{i=1}^N \mu_{i,j} X_i}{\sum_{i=1}^N \mu_{i,j}}$

 Rather than the mean (which likely doesn't correspond exactly to any data point), have the cluster center be the data point closest to the mean

Regression-based modeling

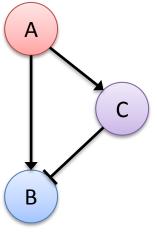
 Relevant if you assume a number of variables (e.g. transcription factors) have independent linear effects

$$Y_g = \sum_{t \in T_g} \beta_{t,g} X_t + \mathcal{E} \longleftarrow \underset{\text{noise}}{\text{Expression of}}$$
 Expression of target gene g on target gene g of TF t

- $\beta_{t,g}$ >0: Transcription factor t positively regulates gene g
- $\beta_{t,g}$ <0: Transcription factor t negatively regulates gene g
- Often, we only want to consider TFs with a large impact on gene expression definitely above noise, so we set a minimum threshold for β or maximum number of nonzero β (other shrinkage methods possible)

Nonlinear effects on gene expression

- Mutual information between pairs of gene expression measurements can detect complex, nonlinear regulatory relationships
 - Feed forward loops



 Cooperativity (multiple subunits to dimerize or multimerize before functional activity)

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