

Your Name: \_\_\_\_\_ TA: \_\_\_\_\_

# 7.012 Quiz 2 Answers

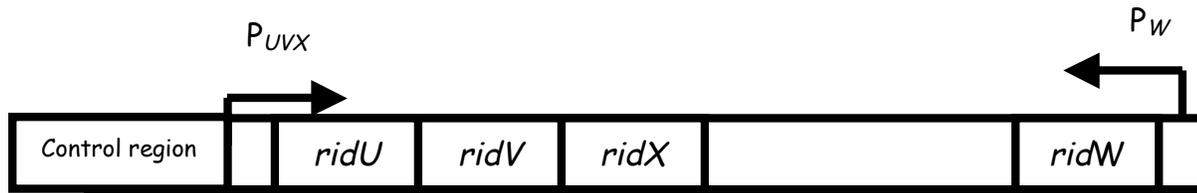
<b>A ≥ 85</b>	<b>~12% of test takers</b>
<b>B ≥ 72</b>	<b>~31.2% of test takers</b>
<b>C ≥ 60</b>	<b>~34.1% of test takers</b>
<b>D ≥ 50</b>	<b>~16.3% of test takers</b>
<b>F ≥ 49</b>	<b>~6.2% of test takers</b>

Regrade requests (with a note attached indicating the problem and part you want looked at) accepted until Thursday November 4<sup>th</sup>, 5pm.

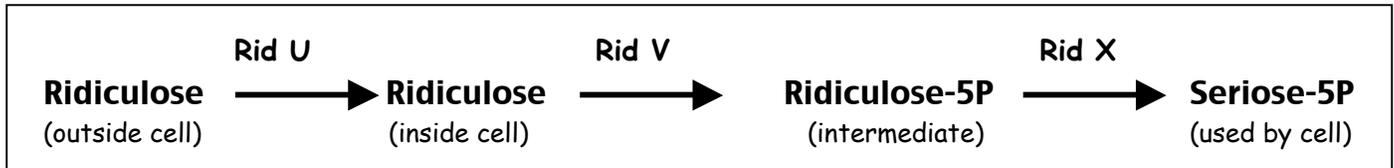
Question	Value	Score
<b>1</b>	<b>17</b>	
<b>2</b>	<b>16</b>	
<b>3</b>	<b>30</b>	
<b>4</b>	<b>17</b>	
<b>5</b>	<b>20</b>	
	<b>100</b>	

### Question 1

In the bacterium *Funditus fabricatus*, the metabolism of the sugar ridiculose is dependent on the *rid* operon shown below.



The ridiculose operon encodes the enzymes shown in the following pathway.



The *ridW* gene is constitutively expressed. The expression of *ridU*, *ridV*, and *ridX* genes is off in the absence of ridiculose and is **activated** by the product of the *ridW* gene in the presence of ridiculose.

There is an artificial inducer of *ridUVX* expression, called *GIG-L*, and an artificial substrate for Rid X, called *STRN* that turns **red** in the presence of active Rid X protein.

a) Several specific Rid- mutants of *F. fabricatus* are shown below. Predict their phenotypes when grown in the presence of *STRN*, with and without the addition of the inducer, *GIG-L*. **10 pts**  
(Fill in the chart with either RED or WHITE)

Strain of <i>F. Fabricatus</i>	+ <i>GIG-L</i>	- <i>GIG-L</i>
WT	RED	WHITE
M1 (deletion of <i>P<sub>UVX</sub></i> )	White	White
M2 (control region that can't bind RidW protein)	White	White
M3 (RidW protein that can't bind ridiculose)	White	White
M4 (nonsense mutation early in <i>ridX</i> )	White	White
M1337 (RidW protein that always binds control region)	Red	Red

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b) Predict whether the following *F. fabricatus* strains, that are merodiploid for the ridiculose operon, will grow on minimal media with or without ridiculose as the **only** carbon source.

Fill in the chart with **YES** if the merodiploid will *GROW* or

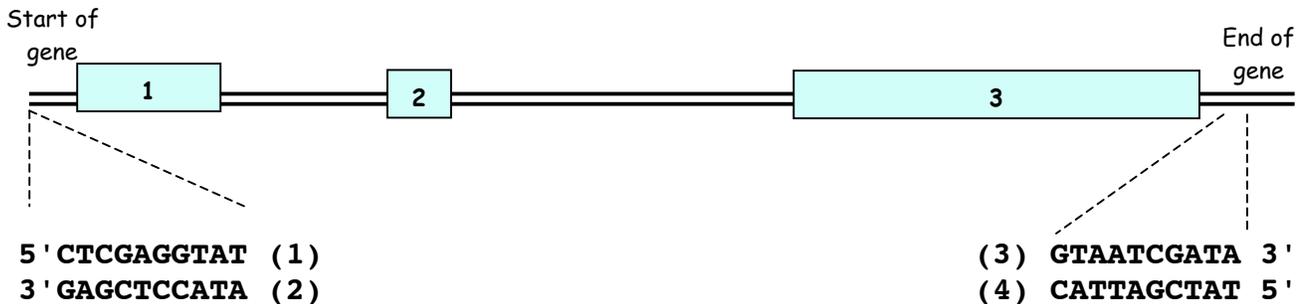
**NO** if the merodiploid will *NOT GROW*

7 pts (5 points for the first column, 2 pts for entire last column)

Merodiploid	+Ridiculose	- Ridiculose
WT / WT	YES	NO
M1 / M2 (deletion of $P_{UVX}$ ) (control region that can't bind RidW)	NO	NO
M2/ M3 (control region that can't bind RidW) (RidW protein that can't bind ridiculose)	YES	NO
M3 / M1337 (RidW protein that can't bind ridiculose) (RidW protein that always binds control region)	YES	NO
M4 / M 1 (nonsense mutation early in <i>ridX</i> ) (deletion of $P_{UVX}$ )	NO	NO
M1337 / M4 (RidW protein that always binds control region) (nonsense mutation early in <i>ridX</i> )	YES	NO

## Question 2

You believe that a disruption in a gene, *sokS*, may contribute to an interesting disease phenotype in cardinals. You wish to PCR amplify and sequence *sokS* from both wild type and diseased cardinals. The *sokS* gene is shown below. Exons are represented by numbered boxes and the terminal sequences are depicted in bold.



a) For PCR amplification, the primers should be identical to which of the following sequences? (Circle one.) 3 pts

1 and 3

2 and 4

1 and 4

2 and 3

b) Which primer(s) should you use in **one** sequencing reaction to obtain sequence that looks most similar to the mRNA (the coding strand)? 2 pts

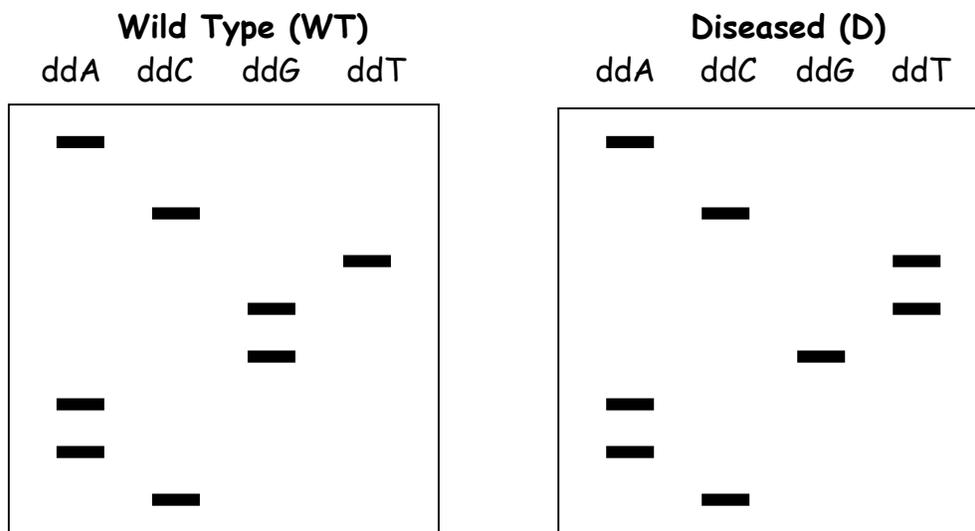
1

2

3

4

You use the dideoxy sequencing method to sequence your PCR products. You see the following pattern in the sequencing gel representing the sequence spanning the end of the first intron and the beginning of exon 2.



c) These gels correspond to which WT and diseased sequences respectively? (WT, D) (Circle i, ii, iii, or iv.) 4 pts

i) 5'-ACTGGAAC-3', 5'-ACTTGAAC-3'

ii) 5'-TGACCTTG-3', 5'-TGAACCTG-3'

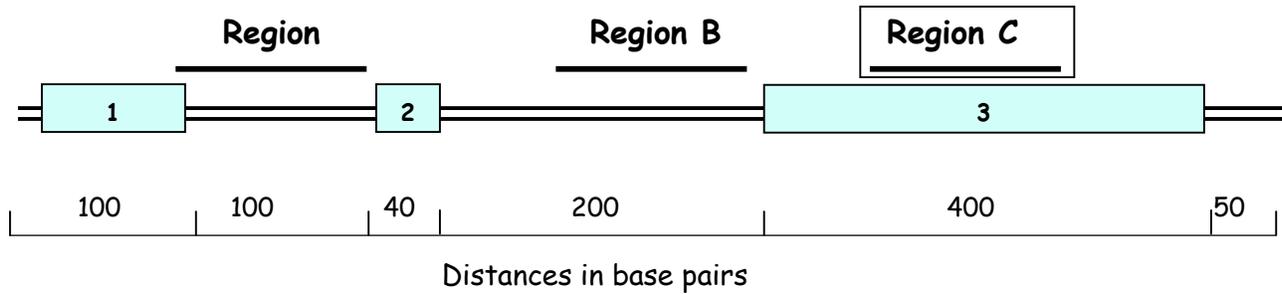
iii) 5'-CAAGGTCA-3', 5'-CAAGTTCA-3'

iv) 5'-GTTCCAGT-3', 5'-5'-GTTCAAGT-3'

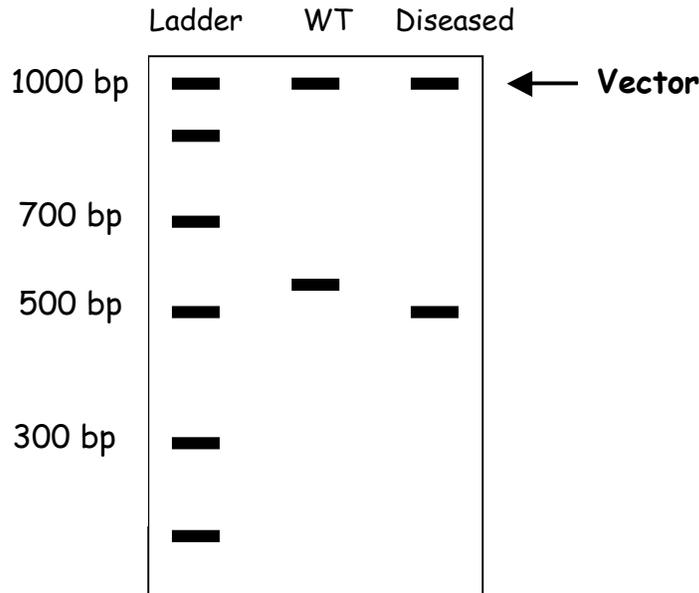
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To determine how this mutated sequence might affect the mRNA and the protein product, you obtain two cDNA libraries: one derived from wild type cardinal RNA and one derived from diseased cardinal RNA.

d) You choose to use a radioactively labeled DNA probe to screen these cDNA libraries for clones containing *sokS*. Which region of *sokS* would make the **best** probe for screening the available cDNA libraries? (Circle either Region A, Region B, or Region C.) 4 pts



Your probe hybridizes to cDNA clones in both cDNA libraries. You purify the plasmids from single clones and cut them with the restriction enzyme used for cloning to verify insert size. The gel is shown below.



e) Based on all evidence above, what is the **most likely** explanation for the difference in restriction enzyme digestion patterns of the *sokS* cDNA clones? 3 pts

i) A mutation in the *sokS* cDNA from the diseased cardinal cDNA library gives rise to an additional restriction enzyme site.

ii) The mRNA encoded by the *sokS* gene from the diseased cardinal is incorrectly spliced.

iii) There is likely a problem with the gel and it should be rerun.

iv) The *sokS* gene from the diseased cardinal acquired a spontaneous insertion.

v) The mRNA encoded by the *sokS* gene from the diseased cardinal has no poly A tail.

Question 3

a) Match the following. Choose **only one** answer for each blank below. 10 pts

- A. Unwinds DNA
- B. Where Okazaki fragments are synthesized
- C. Where DNA can be replicated continuously
- D. Synthesizes RNA primers on DNA
- E. Synthesizes DNA primers on RNA
- F. Catalyzes the addition of dNTPs to lipids
- G. Relieves tension in DNA caused by unwinding
- H. 5' to 3' proofreading activity
- I. 3' to 5' proofreading activity

\_\_B\_\_ lagging strand

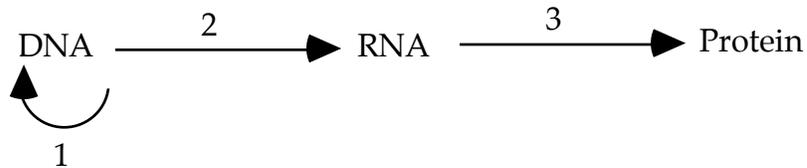
\_\_I\_\_ DNA polymerase

\_\_G\_\_ topoisomerase

\_\_D\_\_ primase

\_\_A\_\_ helicase

b) Where in the eukaryotic cell does the following processes of the central dogma occur?  
One word answer for each, and please write legibly. 6 pts



Process 1 nucleus

Process 2 nucleus

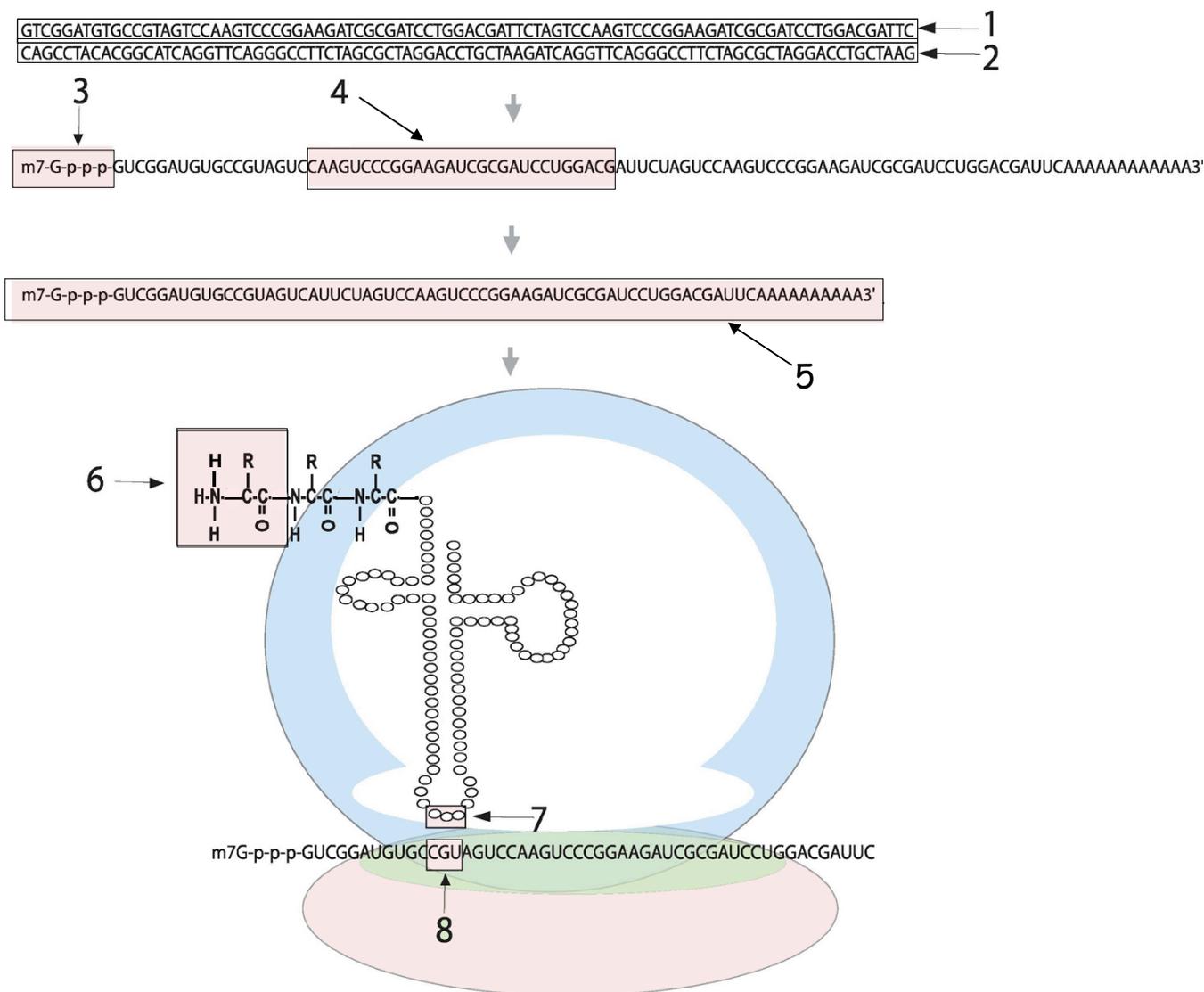
Process 3 cytoplasm

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c) Below are schematics of transcription, splicing and translation. Match the numbered boxes with the following terms. Use each number only once. It's okay to leave blanks.

8 pts

- |                                      |                         |                    |                    |
|--------------------------------------|-------------------------|--------------------|--------------------|
| ___3___ 5' cap                       | ___3' Cap               | ___6___ Amino acid | ___7___ Anti-codon |
| ___1___ Coding (Non-template) strand | ___8___ Codon           | ___ Exon           |                    |
| ___4___ Intron                       | ___5___ mature mRNA     | ___ PolyA tail     |                    |
| ___ Pre-mature mRNA                  | ___2___ Template strand | ___ tRNA           |                    |



d) In the box below, write the sequence that would be in box 7 above. Designate the 5' and 3' orientations. 3 pts

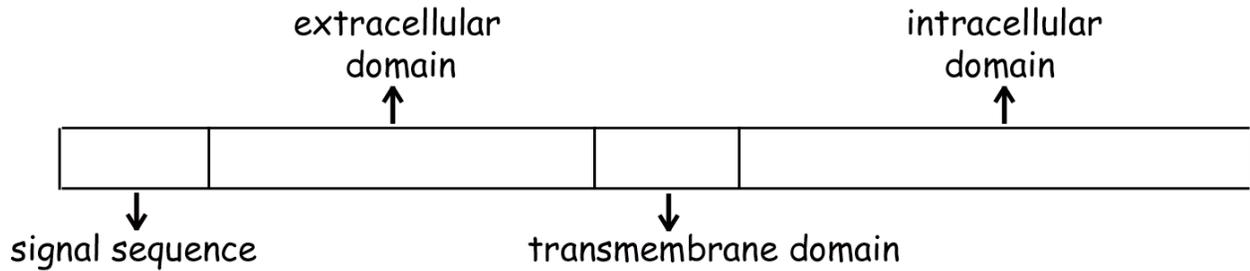
5' - ACG - 3'

e) Circle the **specific** name of the structure that is in box 6 above. 3 pts.

- Alanine    Arginine    Cysteine    Cytosine    Cyanide    Methionine    Threonine    Uracil

Question 4

Below is a diagram of a transmembrane protein called Soxwin.



Normally Soxwin is found embedded in the plasma membrane. You obtain a mutant in which Soxwin is mislocalized. The mutation resides in the DNA that encodes the transmembrane domain.

coding sequence in WT:

...GTTATATTTCTCGTATGGCTCGGCGTCTTATGG...  
 val ile phe leu val trp leu gly val leu trp

coding sequence in mutant:

...GTTATATGTTCTCGTATGGCTCGGCGTCTTATGG...  
 val ile cys ser arg met ala arg arg leu met

a) What type of mutation occurred in the *soxwin* gene? Circle your answer(s). 2pts

deletion     frameshift     insertion    missense    nonsense    silent

b) How has this mutation changed the **chemical property** of the transmembrane domain? Fill in each blank with **one** word. 2pts

From hydrophobic, non-polar in WT to hydrophilic, polar, or charged in mutant

c) Where do you expect the majority of the mutant Soxwin to accumulate? Circle your answer. 3 pts

cytoplasm                      endoplasmic reticulum                      golgi apparatus  
    mitochondria    nucleus                       outside the cell  
 peroxisomes                      plasma membrane                      ribosomes

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d) There's another *soxwin* mutant in which a missense mutation abolishes the function of the signal sequence. Where would you expect the majority of this mutant Soxwin protein to accumulate? Circle your answer. 3 pts

- cytoplasm      endoplasmic reticulum      golgi apparatus  
 mitochondria      nucleus      outside the cell  
 peroxisomes      plasma membrane      ribosomes

e) Match the following. (Multiple answers may be chosen for each blank.)

7 pts

Destinations of proteins

Molecular events

---

\_\_\_C\_\_\_ cytoplasm

A. co-translational transport

\_\_\_B, C\_\_\_ mitochondria

B. post-translational transport

\_\_\_B, C\_\_\_ nucleus

C. entire protein synthesized on a free ribosome

\_\_\_A, D\_\_\_ extracellular space

D. signal sequence recognized by SRP

Question 5

a) A plasmid is a... (Circle your answer(s).) 2pts

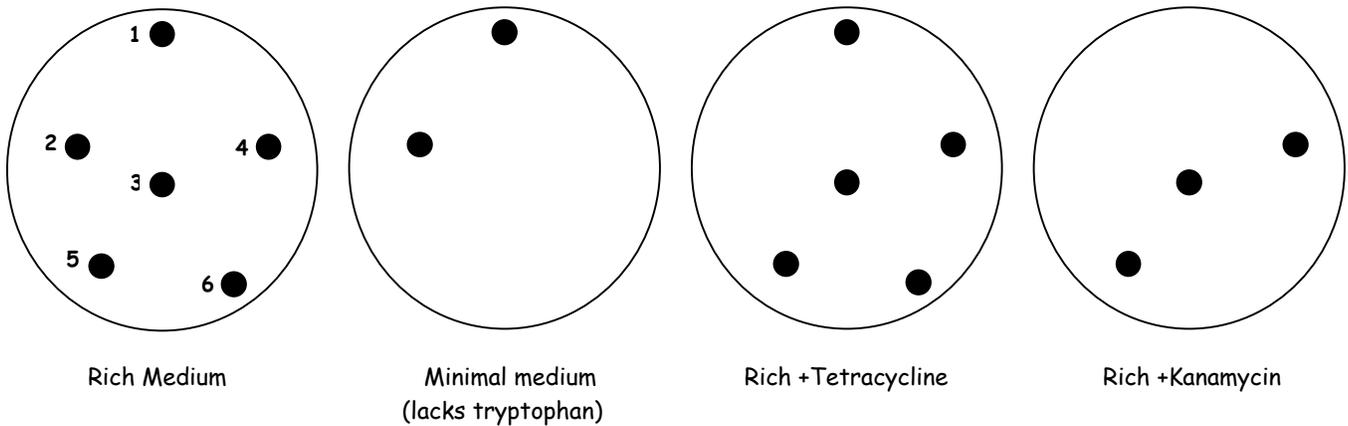
bacterium circular piece of DNA cell multipurpose enzyme petri plate vesicle

b) Match each vector feature with its function. Not all answers need be used. 8 pts

- |                               |  |
|-------------------------------|--|
| ___B___ Restriction site      | A) Required for expression of insert                             |
| ___E___ Origin of replication | B) Allows for insertion of DNA into vector                       |
| ___A___ Promoter              | C) Encodes an enzyme to cut DNA                                  |
| ___D___ Drug resistance       | D) Enables selectability for strain that has taken up the vector |
|                               | E) Required for duplication of vector                            |
|                               | F) Required for SRP to bind                                      |
|                               | G) Site for ribosome to bind                                     |

c) To obtain the gene that rescues a tryptophan biosynthesis *E.coli* mutant strain named, NY-*trp*-Zup, you construct a genomic library from wild-type *E.coli* by cutting the genome with *Bam*HI and inserting the fragments into *pGoSOX!*, a plasmid which has been very successful in the lab. *pGoSox!* contains the genes for tetracycline and kanamycin resistances and has a unique *Bam* HI restriction site that maps to the kanamycin resistance gene. You transform the library into NY-*trp*-Zup and plate the transformants onto rich agar medium. You replica plate the colonies onto different media shown below.

Below are the plates shown in the same orientation after colonies form.



i) Which colony (ies) contain the **original** *pGoSOX!*? 3 pts 1@

None      1      2      3      4      5      6

ii) Which colony (ies) carry *pGoSOX!* containing an insert? 4 pts 2@

None      1      2      3      4      5      6

iii) Which colony (ies) would you choose to further study the gene encoding the tryptophan biosynthetic enzyme that is deficient in NY-*trp*-Zup? 3 pts 3@

None      1      2      3      4      5      6



## The Genetic Code

	U	C	A	G	
U	UUU phe (F)	UCU ser (S)	UAU tyr (Y)	UGU cys (C)	U
	UUC phe (F)	UCC ser (S)	UAC tyr (Y)	UGC cys (C)	C
	UUA leu (L)	UCA ser (S)	UAA STOP	UGA STOP	A
	UUG leu (L)	UCG ser (S)	UAG STOP	UGG trp (W)	G
C	CUU leu (L)	CCU pro (P)	CAU his (H)	CGU arg (R)	U
	CUC leu (L)	CCC pro (P)	CAC his (H)	CGC arg (R)	C
	CUA leu (L)	CCA pro (P)	CAA gln (Q)	CGA arg (R)	A
	CUG leu (L)	CCG pro (P)	CAG gln (Q)	CGG arg (R)	G
A	AUU ile (I)	ACU thr (T)	AAU asn (N)	AGU ser (S)	U
	AUC ile (I)	ACC thr (T)	AAC asn (N)	AGC ser (S)	C
	AUA ile (I)	ACA thr (T)	AAA lys (K)	AGA arg (R)	A
	AUG met (M)	ACG thr (T)	AAG lys (K)	AGG arg (R)	G
G	GUU val (V)	GCU ala (A)	GAU asp (D)	GGU gly (G)	U
	GUC val (V)	GCC ala (A)	GAC asp (D)	GGC gly (G)	C
	GUA val (V)	GCA ala (A)	GAA glu (E)	GGA gly (G)	A
	GUG val (V)	GCG ala (A)	GAG glu (E)	GGG gly (G)	G