



Massachusetts Institute of Technology  
 Harvard Medical School  
 Brigham and Women's/Massachusetts General Hosp.  
 VA Boston Healthcare System



2.79J/3.96J/20.441/HST522J

## SURFACE CHARACTERIZATION AND ANALYSIS

M. Spector, Ph.D.

### 10.7 SIZE AND TIME SCALES FOR BIOADHESION

<u>Size Scale</u>	<u>Tissue Level</u>	<u>Mechanism of Bonding</u>	<u>Time Constant</u>	<u>Measurement(s)</u>
mm-cm	Organ	Interference Fit Grouting Agent Tissue (Bone) Ingrowth Chemical Bonding	Weeks- Months-Years	Radiographic (qualitative) Mechanical Testing (quantitative)
mm	Tissue	Same	Weeks	Mechanical Testing Light Microscopy/Histology (qualitative) Scanning Electron Microscopy (qualitative and quantitative)

### 10.6 BIOADHESION (TISSUE BONDING): PHYSICAL AND CHEMICAL MECHANISMS

1. Physical/Mechanical
  - a. Entanglement of macromolecules (nm scale)
  - b. Interdigititation of ECM with surface irregularities/porosity ( $\mu\text{m}$  scale)
2. Chemical
  - a. Primary ionic
  - b. Secondary
    - 1) hydrogen bonding
    - 2) van der Waals
  - c. Hydrophobic Interactions

$\mu\text{m}$	Cell	Integrin	Days-Weeks	Histology Transmission Electron Microscopy (qual.)
nm	Protein GAG	Secondary Bonding Hydrophobic Interactions	Seconds-Minutes- Hours-Days	Immunohistochemistry (qual.) Adsorption Isotherm (quan.)
nm	Mineral crystallites	Epitaxy Ionic Bonding	Seconds-Minutes- Hours-Days	Transmission Electron Microscopy <i>In vitro</i> Precipitation (quan.)

## CHEMICAL BONDING

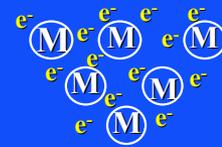
### Primary

- Metallic 100 kcal/mol
- Covalent 200
- Ionic 10-20

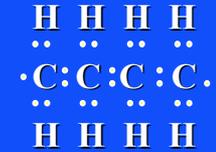
### Secondary

- van der Waals 1-2
- Hydrogen 3-7
- Hydrophobic Interactions 1-2

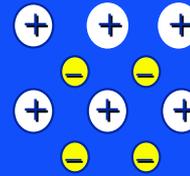
## MATERIALS WITH PRIMARY ATOMIC BONDS



**Metallic**  
(electron "glue"  
or "cloud")  
-metals  
-100 kcal/mol

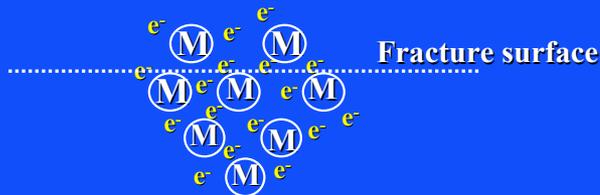


**Covalent**  
(shared- pair electrons)  
-polymers  
-biological macromolec.  
(e.g., proteins)  
-200 kcal/mol

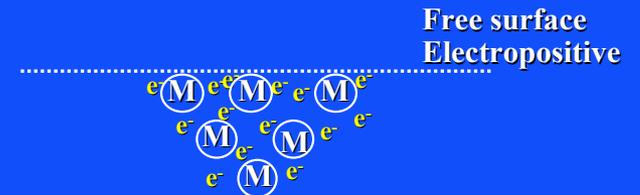


**Ionic**  
(attraction  
of positive  
and negative ions)  
-ceramics  
-calcium phosphates  
-10-20 kcal/mol

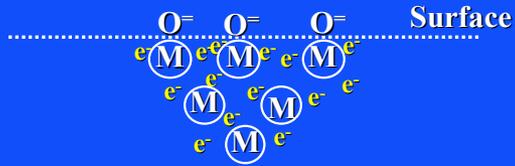
## METALS



## METAL SURFACE



## FORMATION OF METALLIC OXIDE



## ORTHOPAEDIC METALS

	<u>ADVANTAGES</u>	<u>DISADVANTAGES</u>
Stainless Steel	Strength Ease of manuf. Availability	Potential for corrosion High mod. of elasticity
Cobalt-Chromium	Strength Rel. wear resist.	High mod. of elasticity
Titanium	Strength Low modulus Corrosion resist.	Poor wear resistance

## METALS FOR TJA: PAST, PRESENT, AND FUTURE

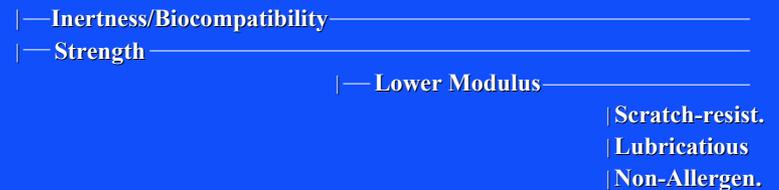


**Oxinium®** (Smith & Nephew Orthopaedics; oxidized zirconium) is the first new metal alloy in orthopaedic surgery in 30 years.

## METALS FOR TJA: PAST, PRESENT, AND FUTURE



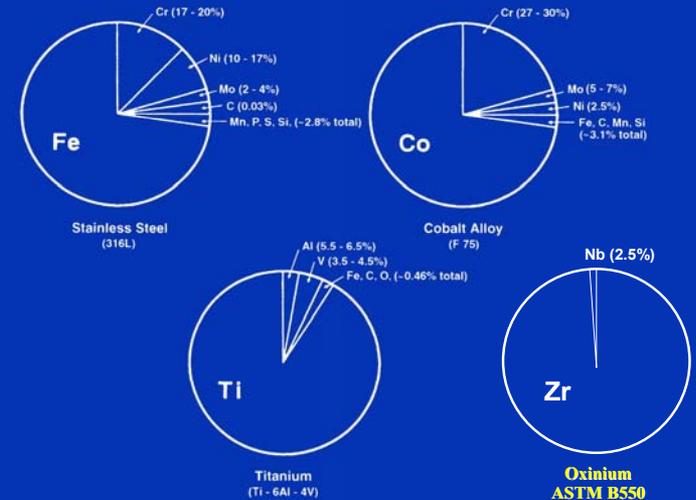
### Selection Criteria



# ORTHOPAEDIC METALS

	<u>ADVANTAGES</u>	<u>DISADVANTAGES</u>
Stainless Steel	Strength Ease of manuf. Availability	Potential for corrosion High mod. of elasticity
Cobalt-Chromium	Strength Rel. wear resist.	High mod. of elasticity
Titanium	Strength Low modulus Corrosion resist.	Poor wear resistance
<b>Oxinium</b>	Scratch-resist. Low modulus	?

## Composition of Orthopaedic Metals



## How is the Ceramic Surface Produced on Oxinium?: Oxidation Process

- Wrought zirconium alloy device is heated in air.
- Metal transforms as oxide grows; not a coating.
- Zirconium Oxide (Zirconia ceramic) is ~5 μm thick.

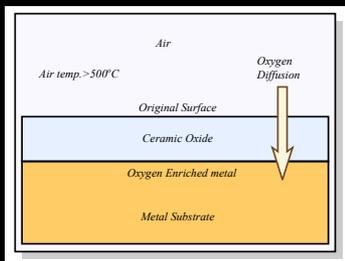
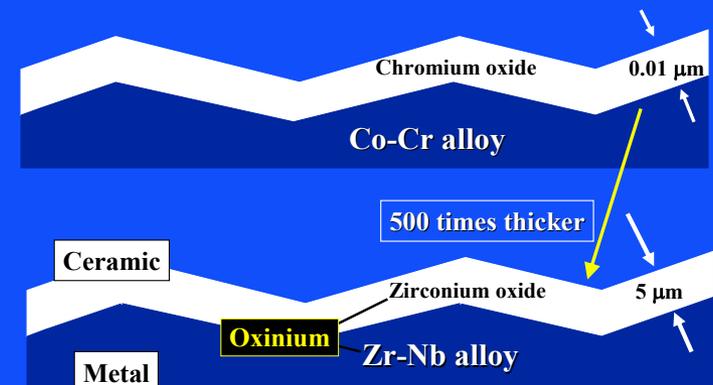
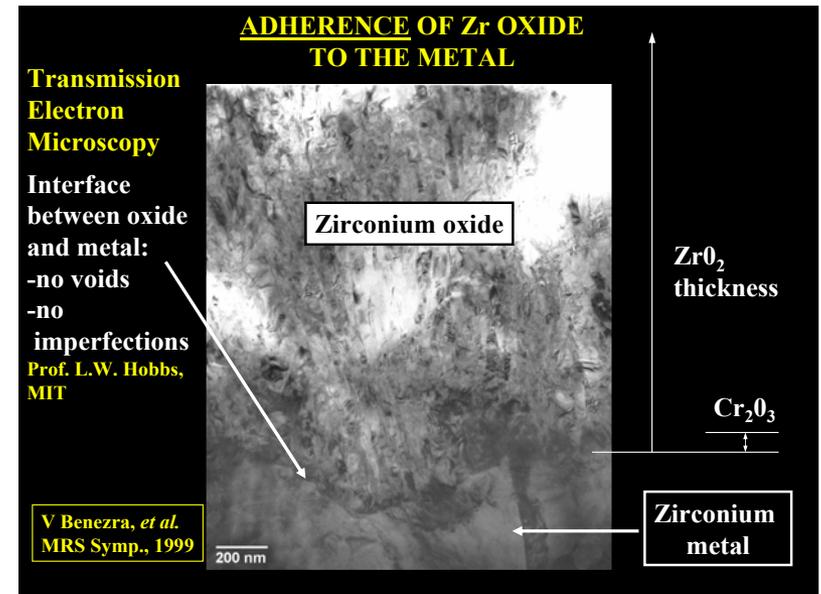
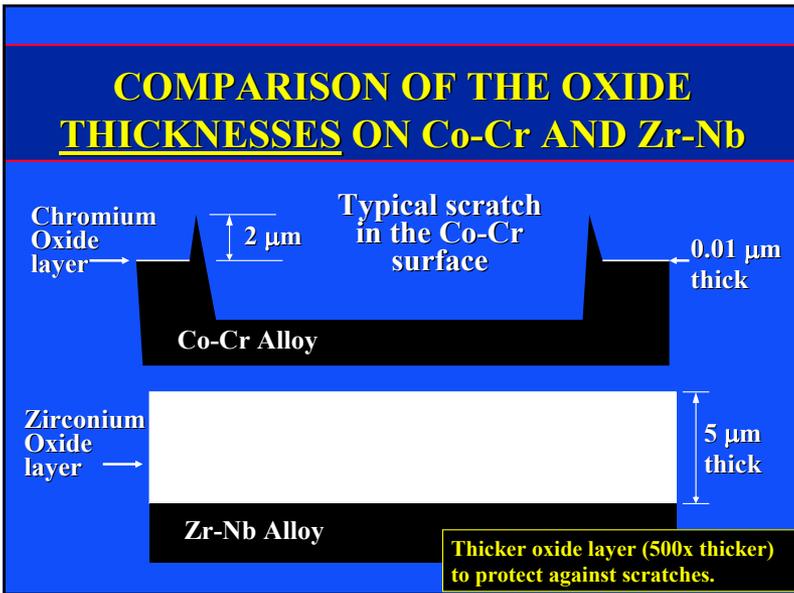


Figure by MIT OpenCourseWare.

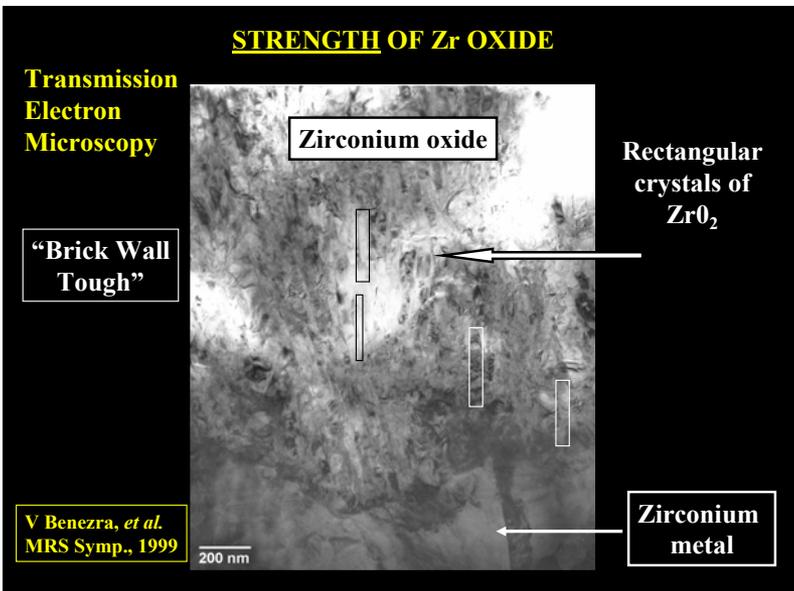
G. Hunter, S&N

## Co-Cr ALLOY VERSUS Zr-Nb ALLOY: THICKNESS OF THE OXIDE

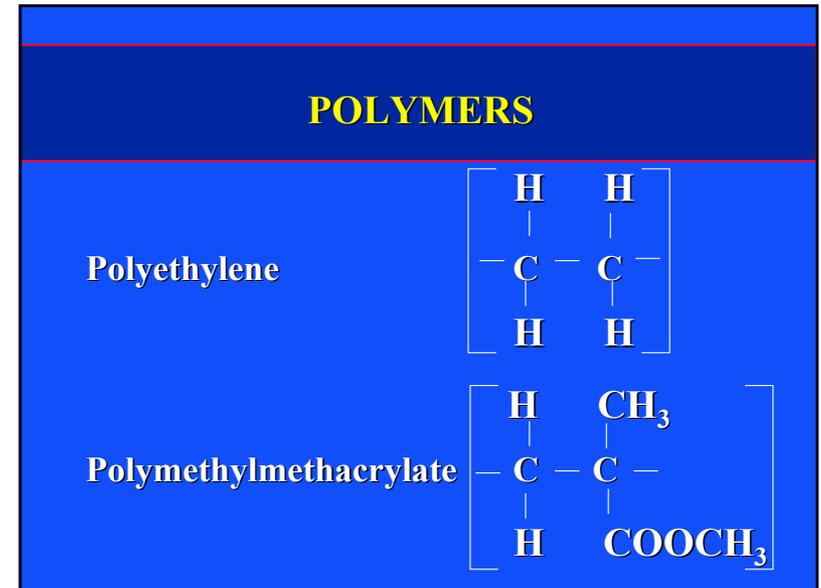




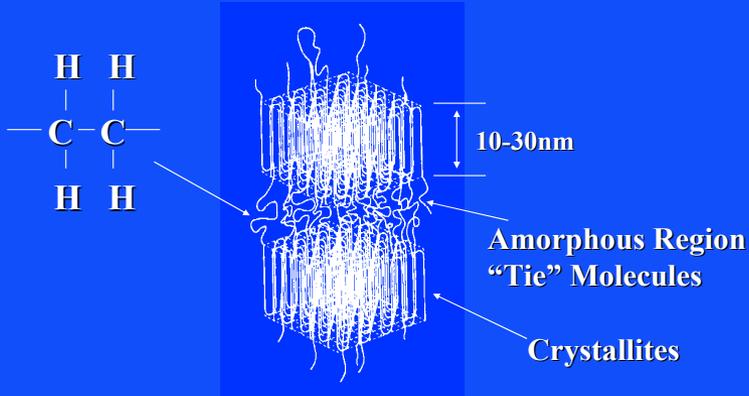
Source: Benezra V., M. Spector et al. "Microstructural investigation of the oxide scale on Zr-2.5 Nb and its interface with the alloy substrate." In: Biomedical Materials -- Drug Delivery, Implants and Tissue Engineering. Mat. Res. Soc. Symp. Proc. Vol. 550 , 1999, pp. 337-342.



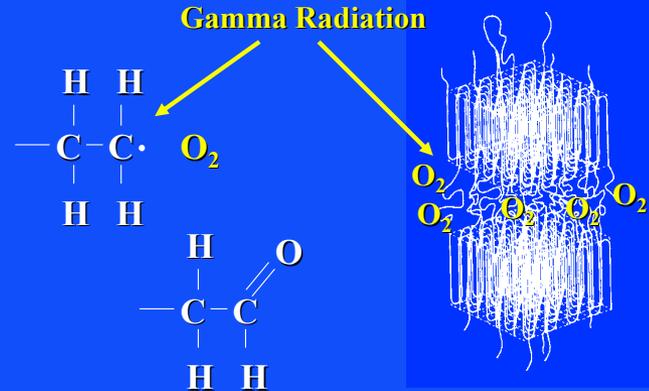
Source: Benezra V., M. Spector et al. "Microstructural investigation of the oxide scale on Zr-2.5 Nb and its interface with the alloy substrate." In: Biomedical Materials -- Drug Delivery, Implants and Tissue Engineering. Mat. Res. Soc. Symp. Proc. Vol. 550 , 1999, pp. 337-342.



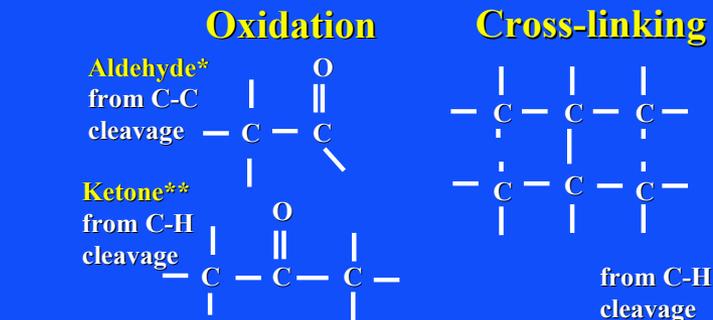
## ULTRAHIGH MOLECULAR WEIGHT POLYETHYLENE



## EFFECT OF GAMMA RADIATION ON PE: OXIDATION



## GAMMA-RADIATION INDUCED MODIFICATION OF POLYETHYLENE



\* Small peak in IR  
\*\* Large peak in IR

Slide removed due to copyright restrictions.  
"Delamination and White Band: Impact of Gamma Sterilization in Air and Material Consolidation."  
Sutula et al, AAOS 1995 Orlando

# CHEMICAL BONDING

## Primary

- Metallic 100 kcal/mol
- Covalent 200
- Ionic 10-20

## Secondary

- van der Waals 1-2
- Hydrogen 3-7
- Hydrophobic Interactions 1-2

## 10.8 CHEMICAL AND PHYSICAL\* BONDING (Nanometer Scale)

Biomaterial			Across Interface	Biological Molecules		
Classification	Bulk	Surface 0.1-5 nm		Intermolecular	Intramolecular	
Metals.....	Metallic	Ionic	Hydrogen (3-7 kcal/mol)	Covalent	Covalent	
Ceramics.....	Ionic/ Covalent	Ionic		van der Waals (1-2)	Ionic	Ionic
Polymers		CE	Ionic** (10-20)		Hydrogen	
- Intramol.....	Covalent			CE		Van der Waals
	.....Ionic	Water (Hydrogel)	Hydrophobic interactions (1-2)			Hydrophobic interactions
- Intermol.....	Covalent					
	..... Ionic					
	..... CE					

\*Physical bonding - chain entanglement (CE), i.e., entanglement of polymer chains with biological macromolecules.

\*\*Includes epitaxial crystal growth of biological mineral (e.g., bone mineral, apatite) on the biomaterial (e.g., synthetic hydroxyapatite or certain metal oxides).

## 10.5 SURFACE CHARACTERISTICS AND METHODS OF ANALYSIS

Scale of Features  
(Not Detection Depth  
of Penetration)

**Macroscopic**  
(>10 μm)

Characteristics	Method
Hydrophobicity	Contact angle (Critical surface tension from Zisman plot)
Charge	Electrophoresis of particles (zeta potential)
Topography	Light microscopy (LM) Scanning electron microscopy (SEM)
Porosity	LM, SEM, Mercury intrusion porosimetry
Water content	Drying/weighing
Surface area	Gas adsorption methods
Mechanical compliance	Mechanical testing (modulus of elasticity)

## Microstructure

(>0.2μm)

Particles on surface	Light microscopy/SEM
Topography Profilometry (stylus pulled)	Light microscopy, SEM, over surface)
Crystallite Structure/Size	X-ray diffraction

<b>Nanostructure</b> >0.01 μm (>10 nm)	Particles	SEM
	Topography	SEM, Profilometry
<b>1-10 nm</b>	Elemental composition analysis (EDX)	Energy dispersive x-ray
		Wavelength dispersive x-ray analysis (WDX) Electron spectroscopy for chemical analysis (ESCA, also referred to as x-ray photoelectron spectroscopy, XPS) Auger electron spectroscopy (AES) Secondary ion mass spectroscopy (SIMS)
	Molecules/Bonding (including depth profile, DP)	ESCA (DP) AES (DP) SIMS (DP) Infrared Spectroscopy (IR)
	Crystal structure	X-ray diffraction (XRD)

## OBJECTIVES OF SURFACE ANALYSIS

- Determine how the surface chemistry (and, therefore, properties) differs from the bulk (relative to the function of the material in the device, effects on the body, and response to effects on the body).
- Identify contaminants (*viz.*, with respect to effects of the material on the body).
- Identify chemical bonding possibilities for interactions with molecules in the biological milieu with respect to the effects of the material on the body (*viz.*, bioadhesion) and the body on the material.

Image removed due to copyright restrictions.  
Comparing visible length scales of unaided human eye, light microscope and electron microscope.

## LIGHT MICROSCOPY

Image removed due to copyright restrictions.  
Diagrams of diascopic and episcopic microscopes.

Slide removed due to copyright restrictions.  
 Description and diagram of compound light microscope.

## LIGHT MICROSCOPY

The resolution (lateral) of the light microscope is:

$$D = \frac{0.611}{N \sin \alpha}$$

$D$  = Smallest lateral dimension that can be resolved

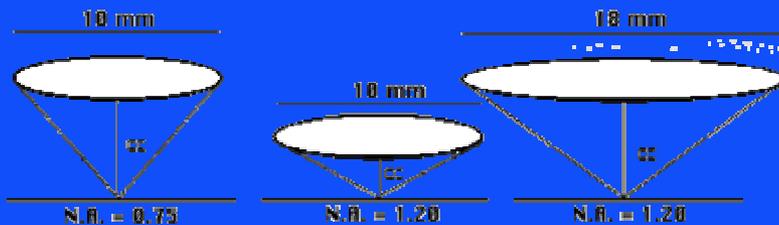
$N$  = Refractive index of medium surrounding the specimen (*i.e.*, air, 1.0, or oil, 1.5)

$\alpha$  = Angular aperture =  $\frac{1}{2}$  angle of cone of light entering the objective lens from the specimen (depends on the width of the objective lens and distance from the specimen) -- increased by moving lens close to the specimen

$N \sin \alpha$  = Numerical aperture

## LIGHT MICROSCOPY

$N \sin \alpha$  = Numerical aperture



## LIGHT MICROSCOPY

For specimens in air viewed by visible light:

$$N = 1.0$$

$$\lambda = 450 \text{ nm}$$

$$D = 292 \text{ (} 0.3 \mu\text{m)}$$

For specimens in oil

$$D = 200 \text{ nm (} 0.2 \mu\text{m)}$$

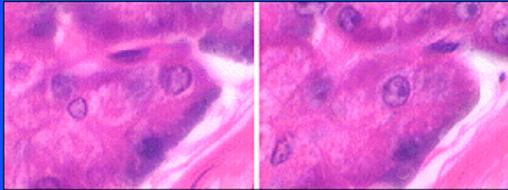
For ultraviolet light  $\lambda = 200 \text{ nm}$

$D$  is approximately  $\frac{1}{2} \lambda$

## LIGHT MICROSCOPY

Another important parameter is depth of focus:

<u>Magnification</u>	<u>Depth of Focus</u>
10X	0.1 mm
100X	1 mm



## Types of Microscopy

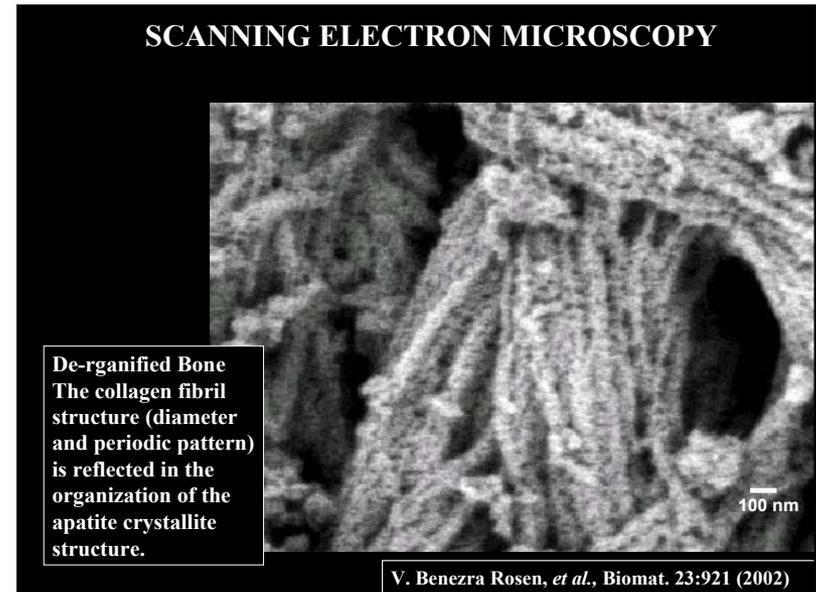
<u>Microscope</u>	<u>Incident Radiation</u>	<u><math>\lambda</math></u>	<u>Resolution (nm)</u>	<u>Depth of Penetration</u>	<u>Depth of Focus</u>
Visible light	Light	450 nm	200	-	1 $\mu\text{m}$ @ 100X
Ultraviolet light	UV	200 nm	100	-	
Electron <sup>1</sup>	e <sup>-</sup>	0.005 (at 50 kV)			
Scanning <sup>2</sup>			2	1 $\mu\text{m}$	1 mm @ 100X
Transmission <sup>3</sup>			0.2	0.1 $\mu\text{m}$ (thickness of section)	

1. Specimen exposed to high vacuum
2. Specimen must have a conducting surface or the use of an "environmental" SEM to prevent "charging"
3. Ultra-thin sections (< 100 nm) are required

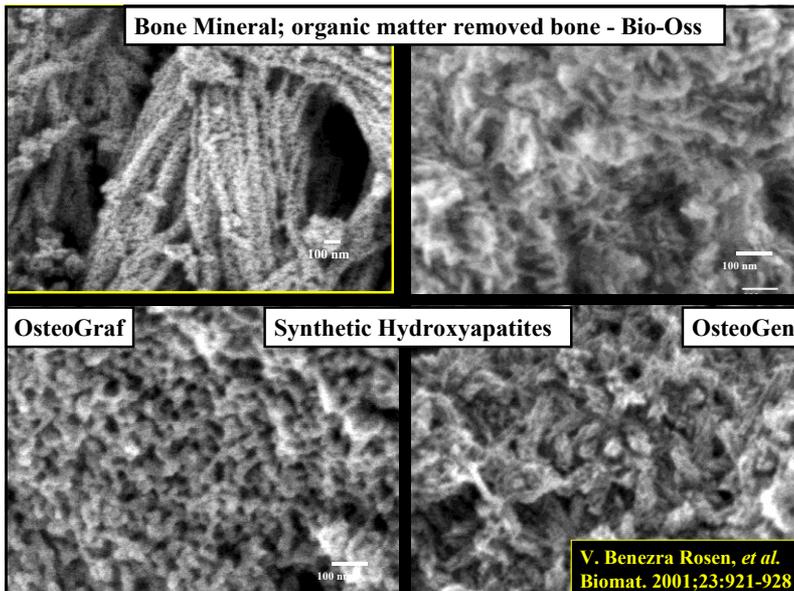
Slide removed due to copyright restrictions.  
Schematic diagram of electron microscope.

Slide removed due to copyright restrictions.  
Interaction with Matter: Secondary electrons interact with topography  
Back scatter electrons interact with composition  
X-rays interact with chemistry

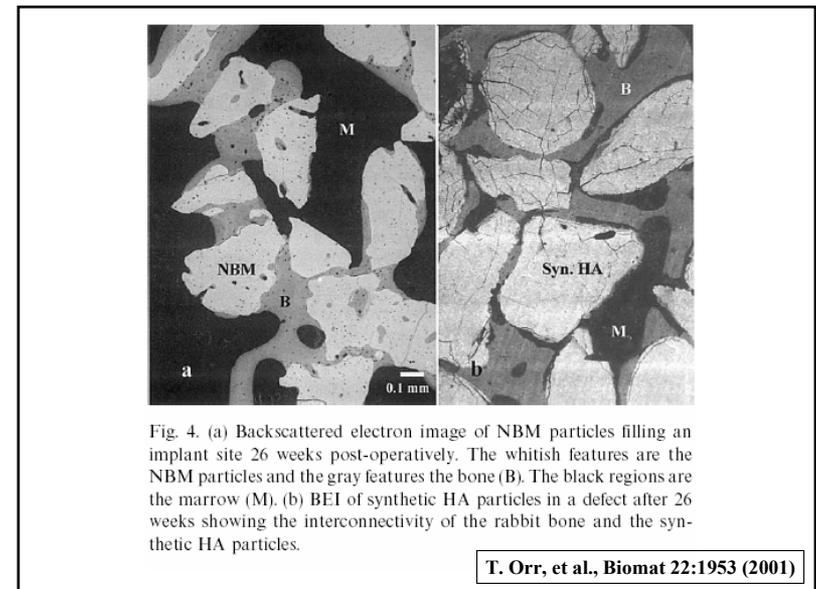
Slide removed due to copyright restrictions.  
 Photos of electron microscope equipment: detectors  
 for secondary and back scatter electrons, and x-rays



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Slide removed due to copyright restrictions.  
Schematic diagram of transmission electron microscope.

Lee DD and Glimcher M, *J. Mol. Bio.* 217:487, 1991  
Lee DD and Glimcher M., *Conn. Tiss. Res.* 21:247, 1989

## TRANSMISSION ELECTRON MICROSCOPY OF BONE

Images removed due to copyright restrictions.

M. Spector, *J Microscopy*  
1975;103:55

60 nm

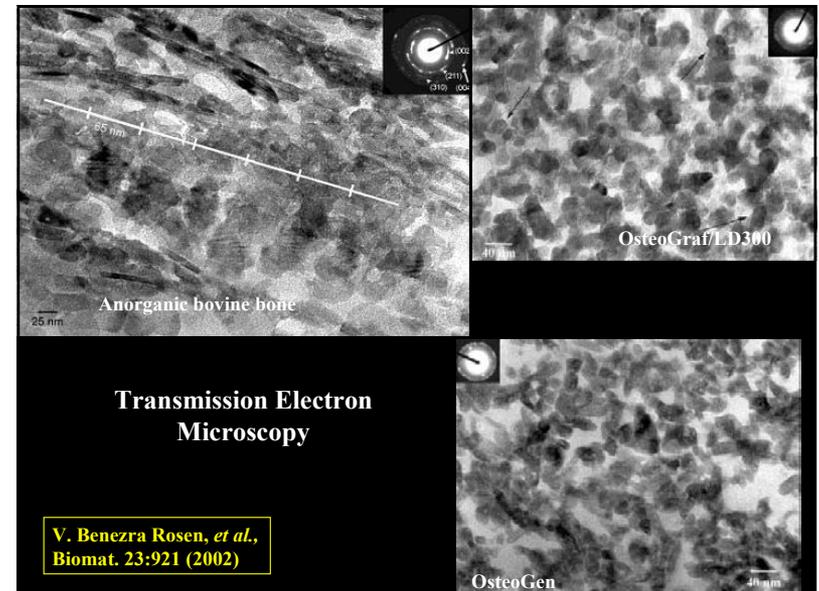
## Transmission Electron Microscopy bONE

Two images removed due to copyright restrictions.  
See Fig 4b and c in Benezra Rosen, V., et al. *Biomat.*  
23:921 (2002).

Whole bone

- Bovine bone from which all the organic matter was removed; anorganic bovine bone; Bio-Oss.
- The crystalline architecture is retained even after removing the organic (collagen) template.

V. Benezra Rosen, *et al.*,  
*Biomat.* 23:921 (2002)



V. Benezra Rosen, *et al.*,  
*Biomat.* 23:921 (2002)

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## Scanning Tunneling Microscopy

Schematic diagram removed due to copyright restrictions.

"Surf. Prop. of Biomat." in *Biomat. Sci.* Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

## Diffraction Methods

- Based on the principle that a monochromatic wave impinging on a **regularly arrayed structure** (e.g., a crystal) will be diffracted at specific angles only, related to the spacing between the features in the array (e.g., molecules).
  - The wavelength of radiation needs to be on the order of (or less than) the spacing to be detected.
- The diffraction pattern is a unique identifying feature of the material.

## Diffraction from a grating

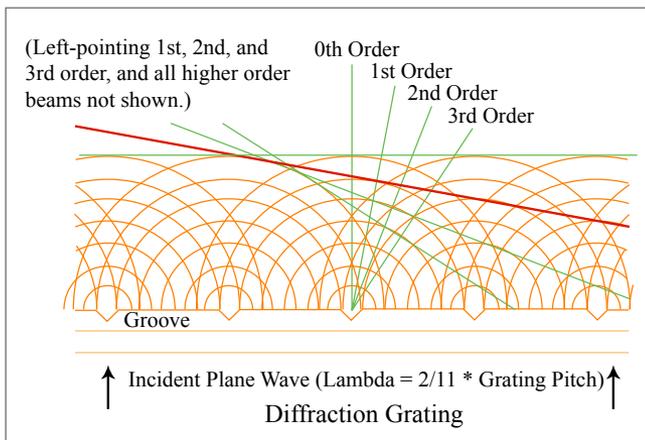
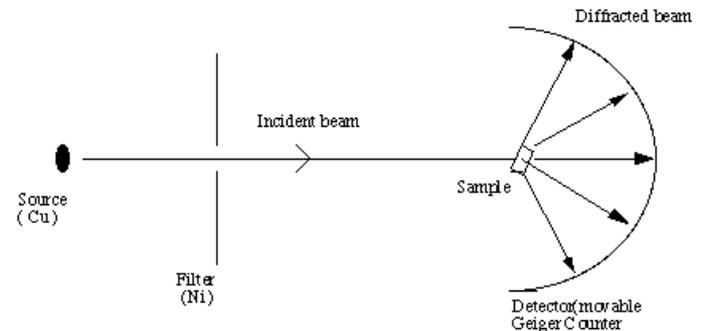


Figure by MIT OpenCourseWare.

## X-RAY DIFFRACTION



**Bragg's Law**

$$n \lambda = 2d \sin \theta$$

$$d = \frac{n \lambda}{2 \sin \theta}$$

$n$  = integer indicating which multiple of the diffracted wave is being considered

$\lambda$  = wavelength of radiation

$d$  = spacing between features in the structure (e.g., spacing between molecules in a crystal)

$\theta$  = the angle between incident and diffracted waves.

Width of peaks related to the crystallite size; narrower peak, larger crystallite

Photo and pair of graphs (natural bone mineral and synthetic hydroxyapatite) removed due to copyright restrictions.

## CRYSTALLOGRAPHIC ANALYSIS (Powder X-ray Diffraction)

Sample	Structural Identification	Crystallinity	Crystallite Size: Peak	Crystallite Size: Size
Anorganic Bone	HA: >99.8% Non HA Ca-P: <0.2%*	88%	(002)	39 nm
Synthetic HA	HA	98%	(002)	90 nm

\* Trace detection of  $\alpha$ - and  $\beta$ -TCP,  $\text{Ca}_2\text{P}_2\text{O}_7$  and CaO was observed however insufficient quantity of material existed for confident quantitative assessment.

START HERE

### Types of Diffraction

Type of Diffraction	Radiation	$\lambda$
Optical	Laser light	400 nm
X- ray	X-ray	0.154 nm (for copper)
Electron	$e^-$	0.005 nm at 50 kV

Depth analyzed for x-ray diffraction (i.e., depth of penetration of the x-ray beam) is 1-10  $\mu\text{m}$ .

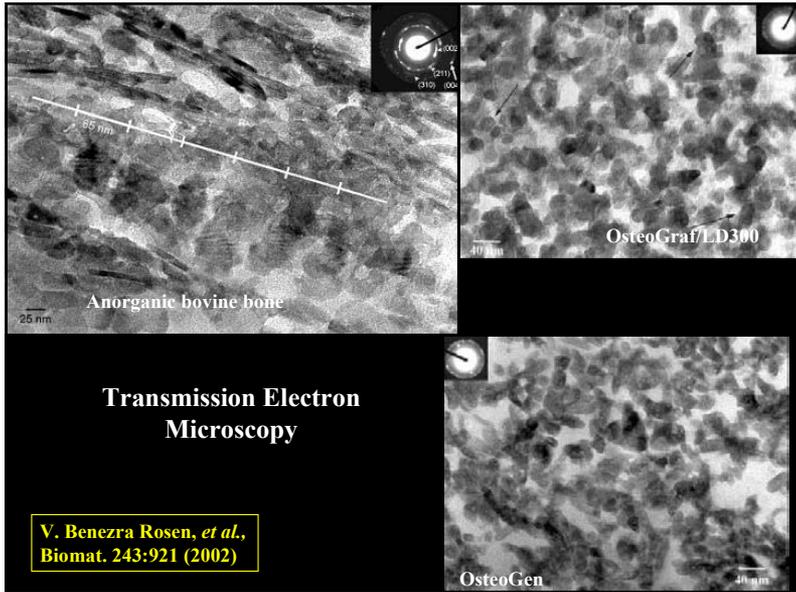


Table 1 (Common Methods of Characterizing Biomaterials Surfaces) from Ratner removed due to copyright restrictions.  
 [Preview in [Google Books](#)]

“Surf. Prop. of Biomat.” in *Biomat. Sci.*, Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

### Contact Angle and Critical Surface Tension

(included angle) = 0: complete wetting  
 0 <  $\theta$  < 90°: partial wetting  
 $\theta$  > 90°: nonwetting

Characterize a surface based on its critical surface tension,  $\gamma_{sv}$ .

At Equilibrium,  $\sum$  surface tensions = 0

$$\gamma_{sv} - \gamma_{sl} - \gamma_{lv} \cos \theta = 0$$

$$\gamma_{sv} - \gamma_{sl} = \gamma_{lv} \cos \theta \quad \text{Young's equation}$$

Cannot solve for  $\gamma_{sv}$  because there are 2 unknowns,  $\gamma_{sl}$  and  $\gamma_{sv}$

The experimental method employed to approximate  $\gamma_{sv}$  involves asking the question:  
 What is the surface tension of a liquid that would completely wet the solid surface?

### Contact Angle Zisman Plot

Graph from Ratner removed due to copyright restrictions.

“Surf. Prop. of Biomat.” in *Biomat. Sci.*, Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

Table 2 (Critical Surface Tension Values for Common Polymers) from Ratner removed due to copyright restrictions.

"Surf. Prop. of Biomat." in Biomat. Sci. Eds., B.D. Ratner, et al., Academic Press, San Diego, CA, 1996

### Contact Angle Assumptions

- **Equilibrium between the liquid droplet and solid surface has been reached (*i.e.*, no absorption of liquid by the solid and no leaking of substances from the solid).**
  - If this assumption cannot be met then the "advancing angle" can be measured to determine the contact angle of the liquid with the dry surface and "receding angle" measured to determine the contact angle with the water absorbed surface.
  - An alternative method is to measure the underwater (captive-air-bubble) contact angle that an air bubble makes with the immersed surface. This is particularly valuable for measuring surface that can switch from hydrophobic to hydrophilic depending on the environment.

### Methods for Measuring the Contact Angle

Figure 5 from Ratner removed due to copyright restrictions.

"Surf. Prop. of Biomat." in Biomat. Sci. Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

Table 3 (Concerns in Contact Angle Measurement) from Ratner removed due to copyright restrictions.

"Surf. Prop. of Biomat." in Biomat. Sci. Eds., B.D. Ratner, et al., Academic Press, San Diego, CA, 1996

## SURFACE ANALYSIS

### Incident Beam

- **X-Ray**
  - X-Ray Photoelectron Spectroscopy
- **Electron**
  - Energy Dispersive X-Ray Microanalysis
  - Auger Spectroscopy
- **Ion Beam**
  - Secondary Ion Mass Spectroscopy
- **Infrared Radiation**
  - Infrared Spectroscopy

### Auger Electron Spectroscopy

- Energy analysis of Auger electrons emitted from a sample. electrons are produced as a result of ionizations in inner core shells under impact of an electron beam.
- Significant intensity occurs for Auger electrons emitted with energies up to 2500 eV and these typically have a characteristic range between 1.5 and about 10 atom layers (0.4 to 3 nm).
- AES is sensitive to about 1% of most elements except H and He in the outermost atom layer and, generally, some of the atom layers just below the surface.
- The excitation is by an electron beam with energies in the range 5 keV to 25 keV.
- These beams may be focused to spot sizes of < 12 nm in most modern instruments.

### X-ray Photoelectron Spectroscopy

#### Electron Spectroscopy for Chemical Analysis (ESCA)

- Energy analysis of photoelectrons emitted from a sample generated from core level shells under impact by characteristic X-rays, usually Al or Mg Ka.
- Photoelectrons have energies up to 1500 eV and typically have a characteristic range between 3 and about 8 atom layers (1 to 3 nm).
- XPS has a similar sensitivity to AES.
- It is sensitive to about 1% of most elements except H and He in the outermost atom layer and, generally, some of the atom layers just below the surface.
- This allows the composition to be determined as a function of depth to 10 nm below the surface, non-destructively.

Figure 7 from Ratner removed due to copyright restrictions.

**Electron Spectroscopy  
for Chemical Analysis,  
ESCA or X-ray  
Photoelectron  
Spectroscopy, XPS**

Figure 6 from Ratner removed due to copyright restrictions.

"Surf. Prop. of Biomat." in *Biomat. Sci.* Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

**ESCA or XPS**

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"Surf. Prop. of Biomat." in *Biomat. Sci.* Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

4 C peaks: hydrocarbon,  
C singly bonded to O, C  
in amide-like  
environment, and C in  
acid or ester  
environments

Figure 9 from Ratner removed due to copyright restrictions.

"Surf. Prop. of Biomat." in *Biomat. Sci.* Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

**SURFACE AND BULK ANALYSIS**

Sample	Analysis Ca/P for§	Composition				Ratio:†
		Ca	P	O	C	
Anorganic	Bulk:	27	16	57	–	Bulk: 1.69
Bone	Surface:	19	13	58	10	Surface: 1.46
Synthetic	Bulk:	25	15	60	–	Bulk: 1.67
Hydroxyapatite	Surface:	20	13	58	9	Surface: 1.54

§ : Analysis for:

Bulk Composition: Energy Dispersive X-ray Analysis (0-2 µm sampling depth)

Surface Composition: Electron Spectroscopy for Chemical Analysis (0-5 nm sampling depth).

† : The stoichiometric ratio of Ca/P for Hydroxyapatite ( $\text{Ca}_{10}(\text{PO}_4)_6(\text{OH})_2$ ) is 1.67.

### Static Secondary Ion Mass Spectrometry

- Mass analysis of positive or negative ions emitted from samples under impact of an energetic ion beam.
- Significant intensity can occur for ions with masses up to and beyond 1000 a.m.u. that originate mainly from the outermost molecular layer.
- The fragments observed in the spectrum reflect the precise molecular groups on the surface.
- This allows distinction and identification not possible by AES and XPS but only where suitable reference spectra are available.
- SSIMS has a very much higher sensitivity than AES or XPS, but quantification is much more complex since matrix effects are dominant.

### Infrared Spectroscopy Attenuated Total Reflectance Mode

Figure 11 A from Ratner removed due to copyright restrictions.

“Surf. Prop. of Biomat.” in Biomat. Sci. Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

### Infrared Spectroscopy External Reflectance Mode

Figure 11 B from Ratner removed due to copyright restrictions.

“Surf. Prop. of Biomat.” in Biomat. Sci. Eds., B.D. Ratner, *et al.*, Academic Press, San Diego, CA, 1996

### Infrared Spectroscopy Diffuse Reflectance Mode

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