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Lecture: “MEMS based sensors for cellular studies” by Dr. Taher Saif.

Given August 10, 2006 during the GEM4 session at MIT in Cambridge, MA.

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# **MEMS based sensors for cellular studies**

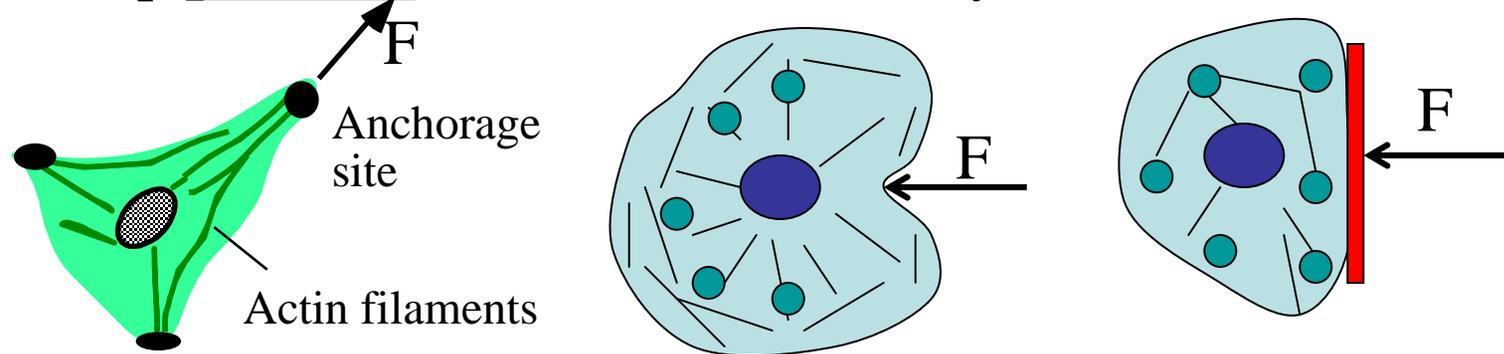
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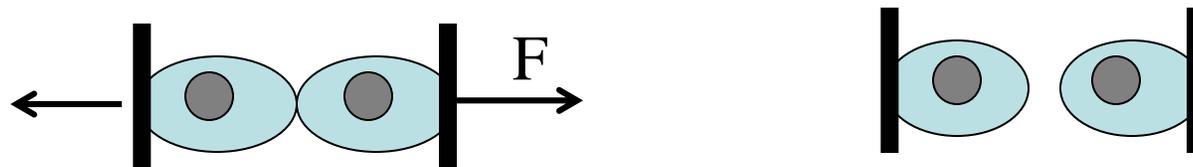
Part of GEM4 Summer School lectures on  
instruments for cell mechanics studies (Aug 10, 2006, MIT)

## Objective

Develop **portable** micro sensors to study:



- Cell mechanical response

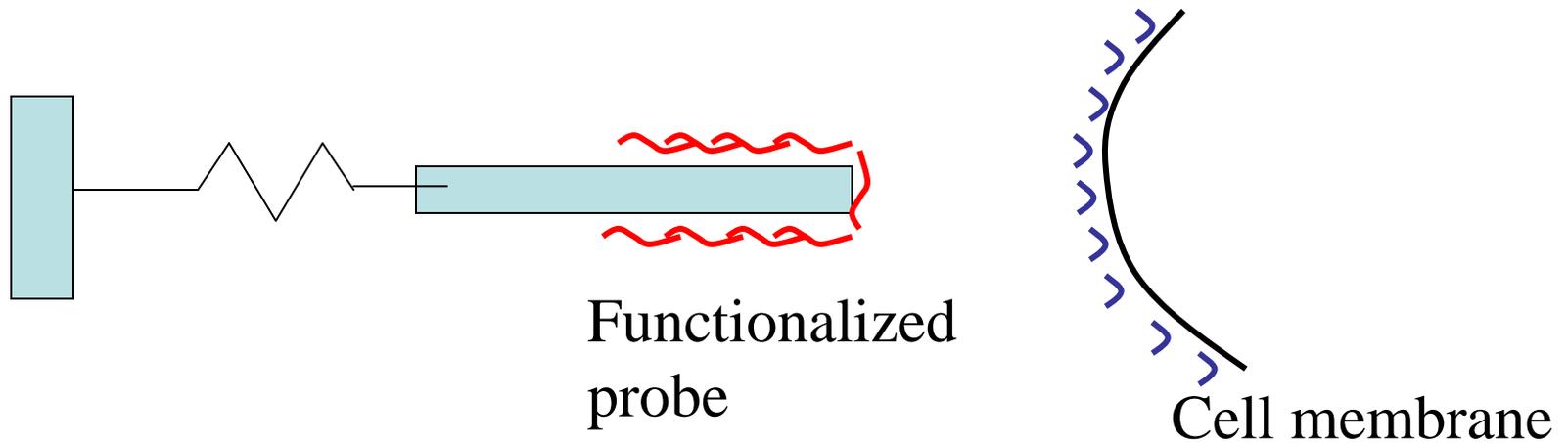


- Cell adhesion

in different biochemical environments  
to explore mechanotransduction and disease detection

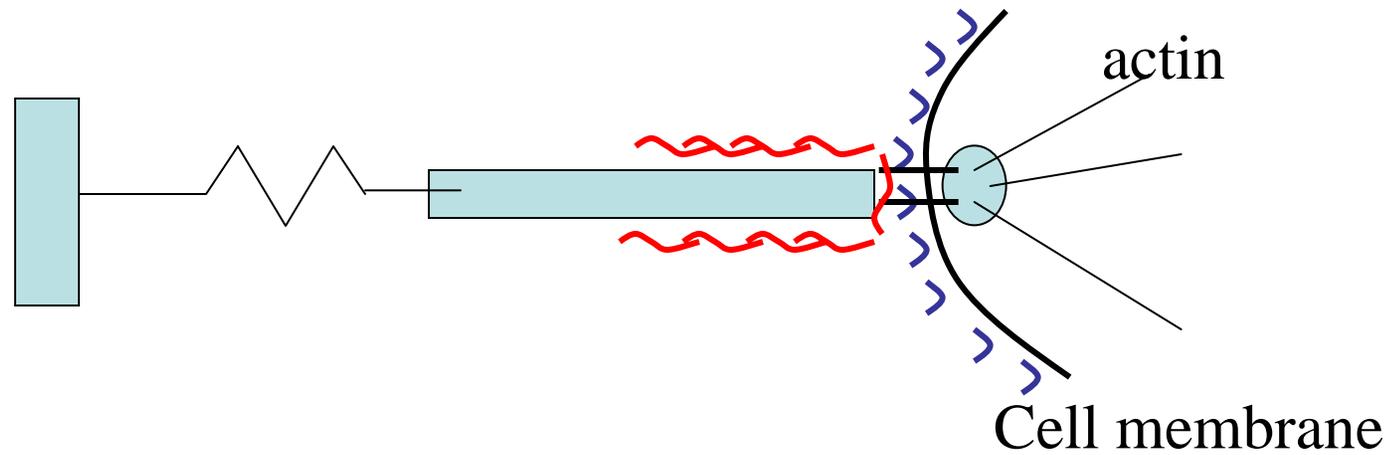
## Basic idea

A micro spring is used to measure cell force



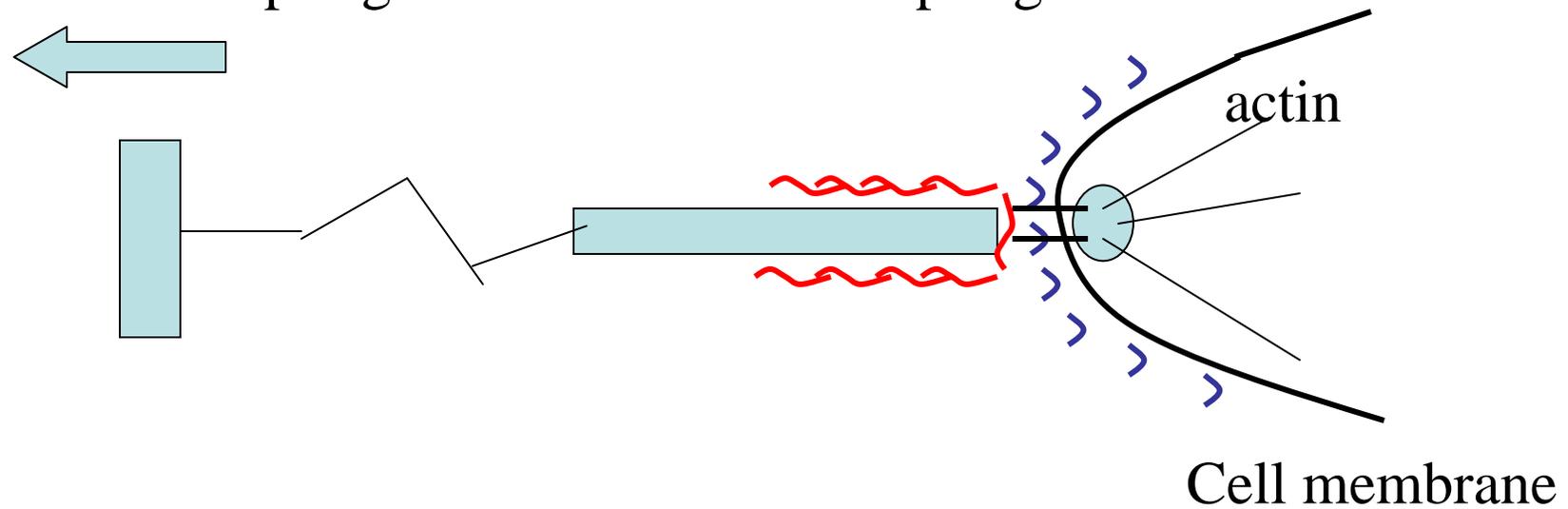
# Basic idea

Functionalized probe contacts a cell and forms adhesion site



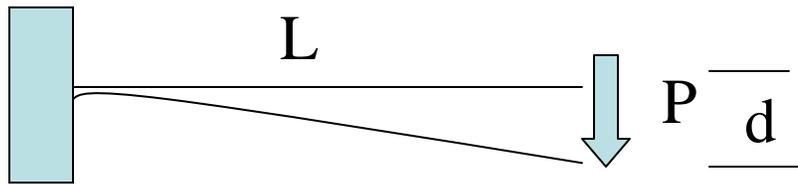
## Basic idea

Probe is moved away from the cell. The cell applies a force on the spring. The force is measured from the spring deformation and its spring constant.



The cell may also be compressed or indented.

# Cantilever as a mechanical spring



$$K \text{ (spring constant) } = 3EI/L^3$$

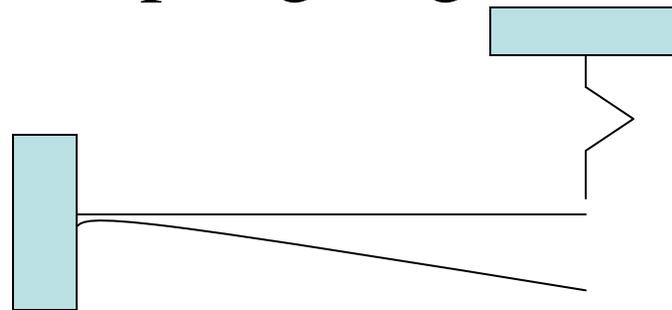
$$P = Kd$$

$I$  = moment of inertia = width x depth<sup>3</sup>/12

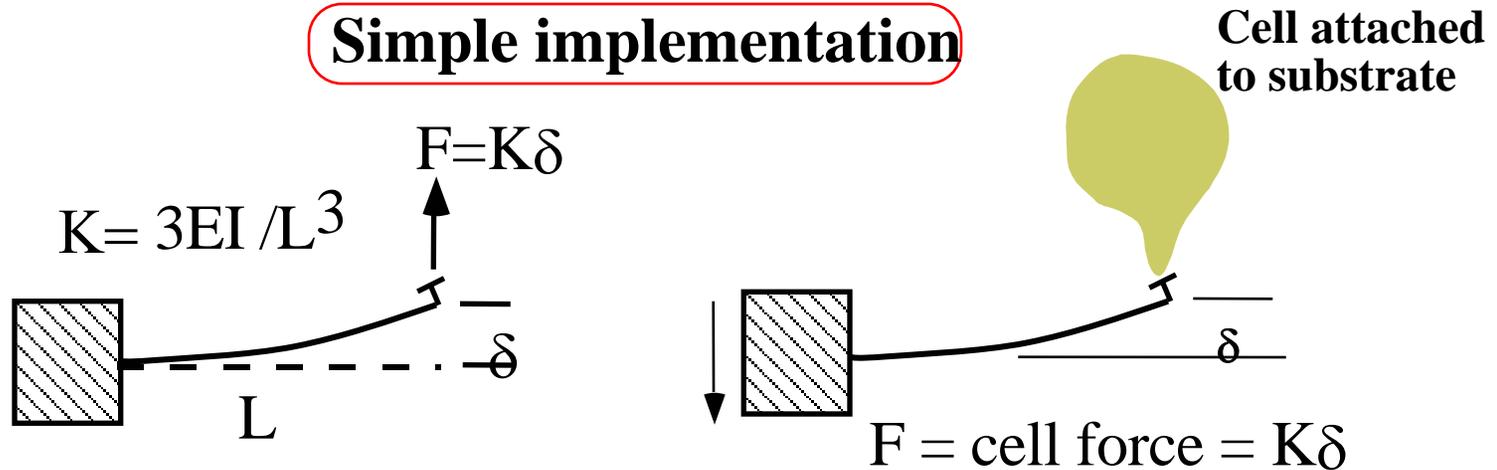
Typical  $K \sim 10 \text{ nN}/\mu\text{m}$

Calibration:

- 1) Resonant frequency, geometry, elastic property
- 2) Comparing with another spring (e.g., AFM)



## Simple implementation

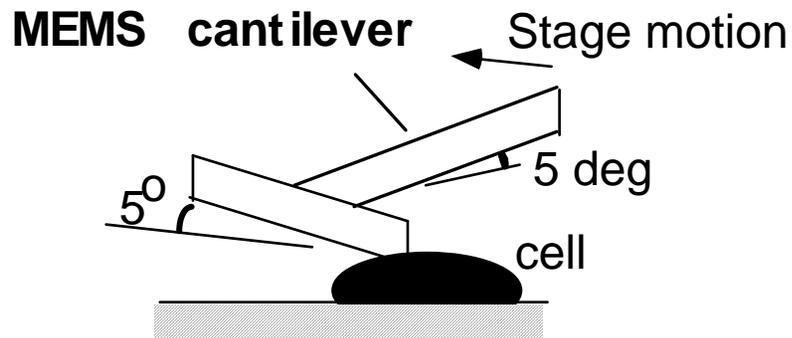
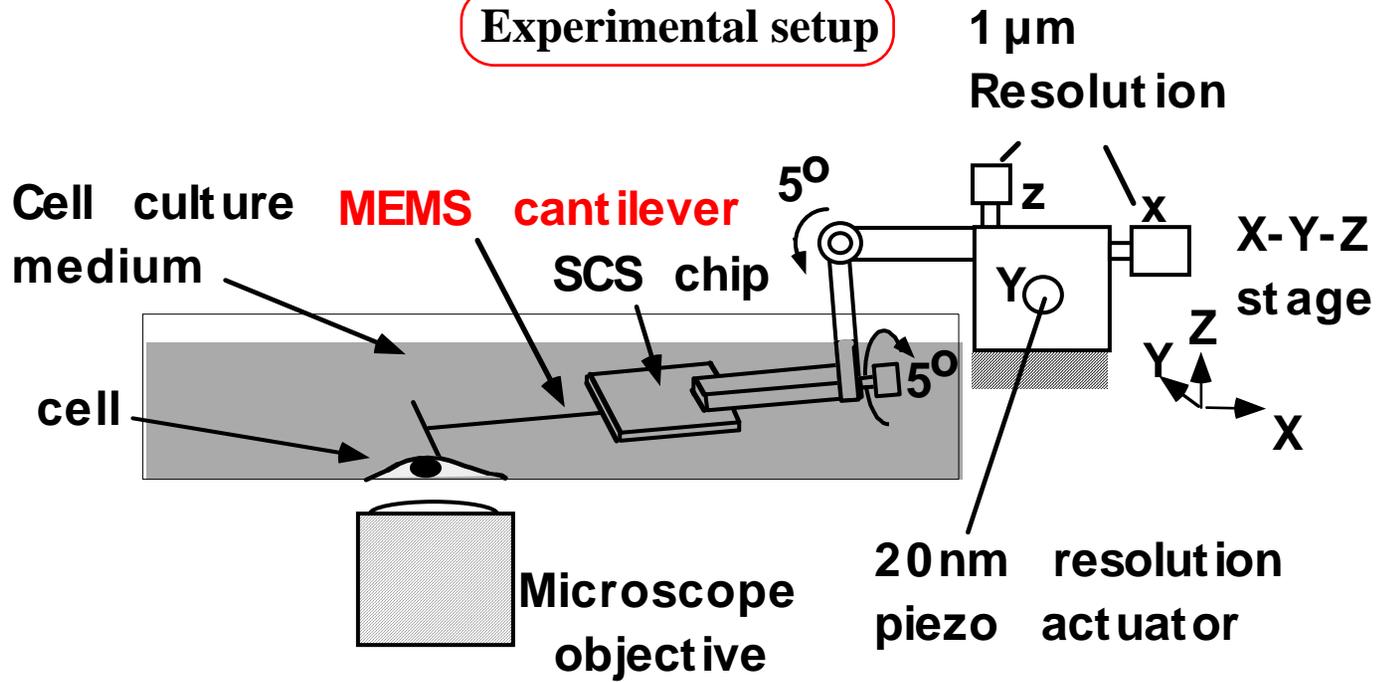


- Force sensor (such as a cantilever) is coated by fibronectin
- It is calibrated to determine spring constant,  $K$
- The sensor tip is brought in contact with the cell - focal adhesion sites form
- It is moved away from the cell. The cell force is measured from the deformation  $\delta$

**Advantages:**

- Force sensor independently calibrated
- Force is applied at an anchorage site
- In-situ observation

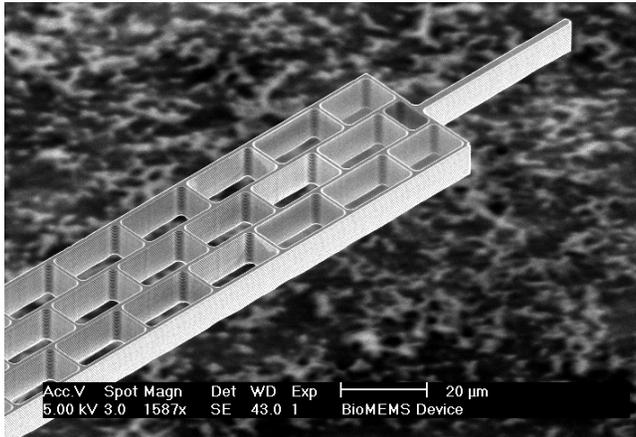
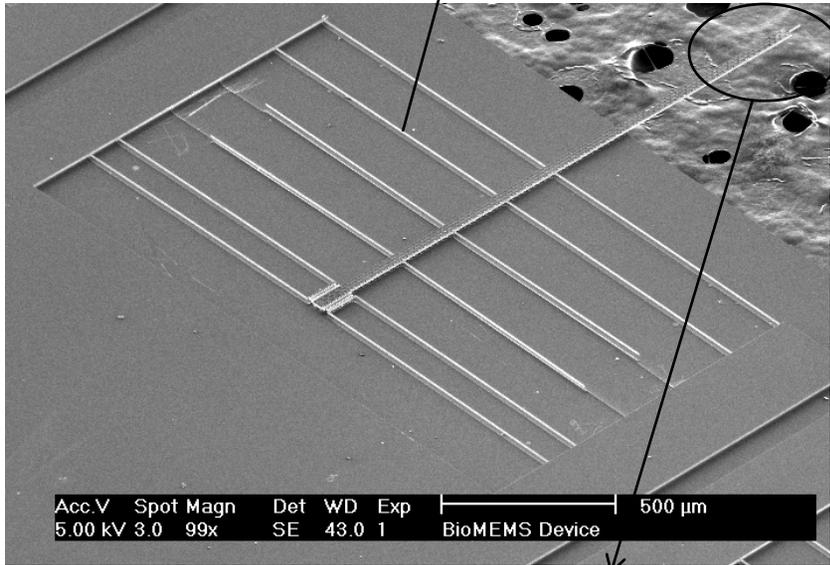
**Experimental setup**



**Example**

**MEMS force sensor: beams anchored at both ends**

**Flexural springs (1 $\mu$ m wide)**



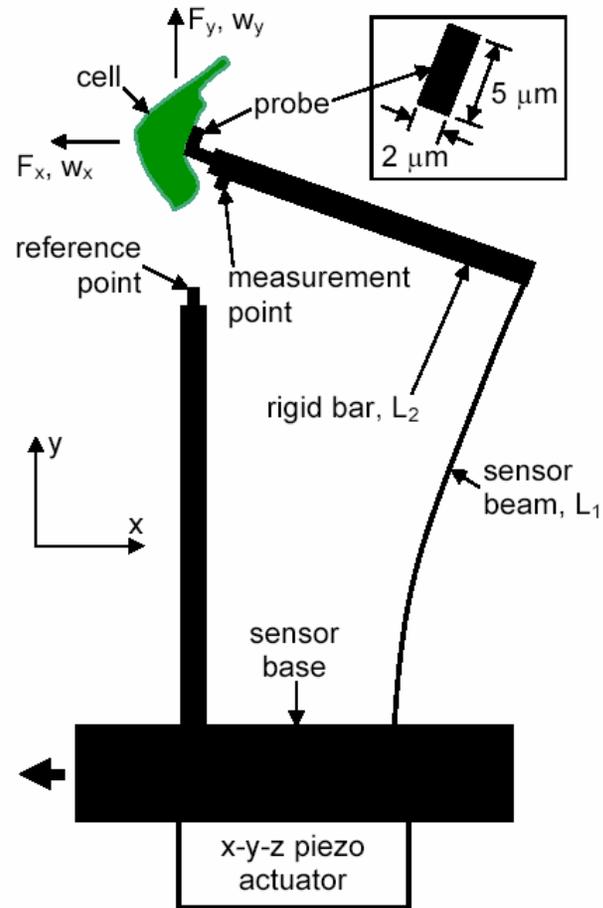
**$K = 3.4 \text{ nN}/\mu\text{m}$**

Yang and Saif.  
Review of Scientific Instruments 76, 044301 (2005).

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**Example**

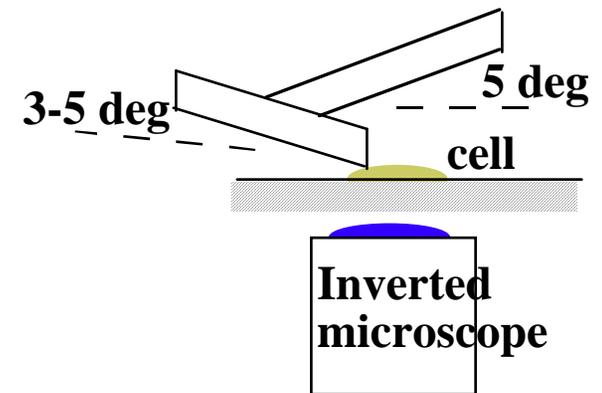
# 2D force sensor



**Example**

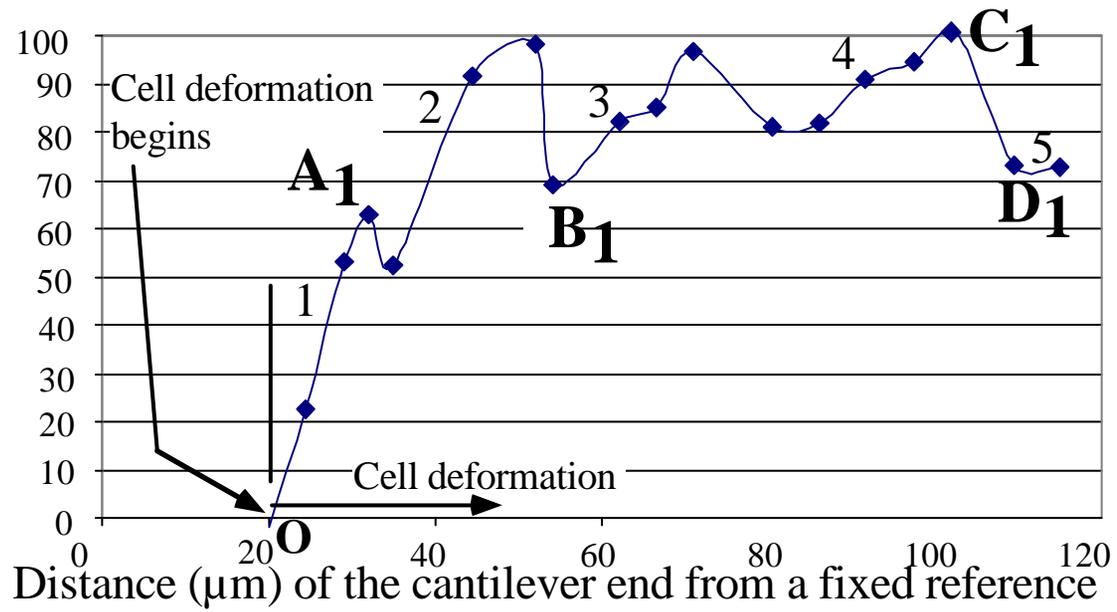
**Endothelial cells in a culture dish with MEMS cantilever**

Image removed due to copyright restrictions.



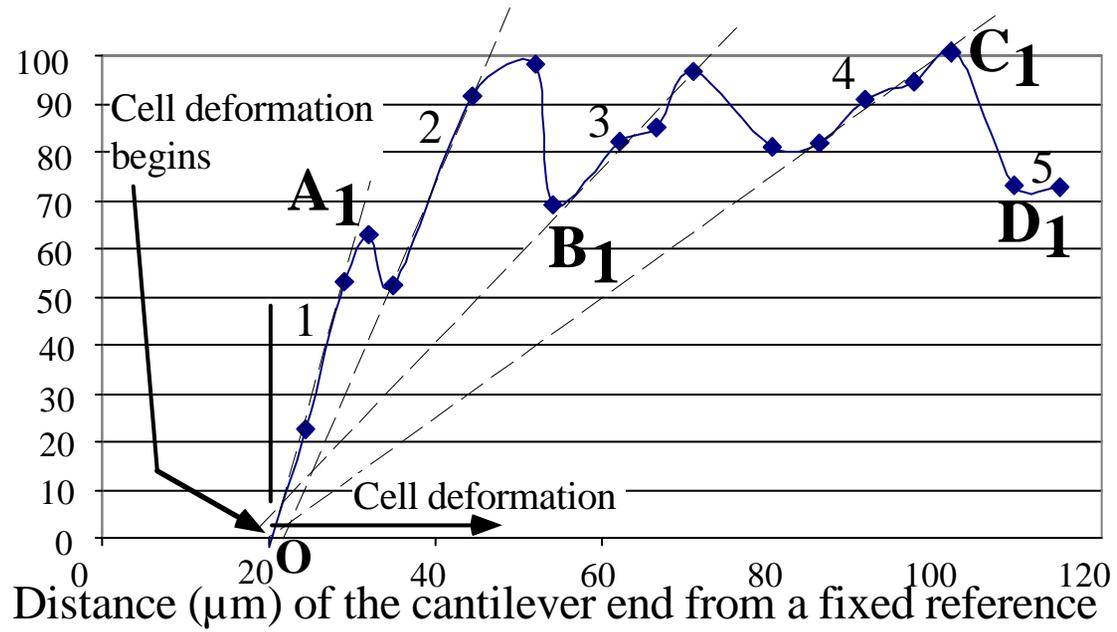
**Schematic of the  
MEMS cantilever**

## Force response of an endothelial cell



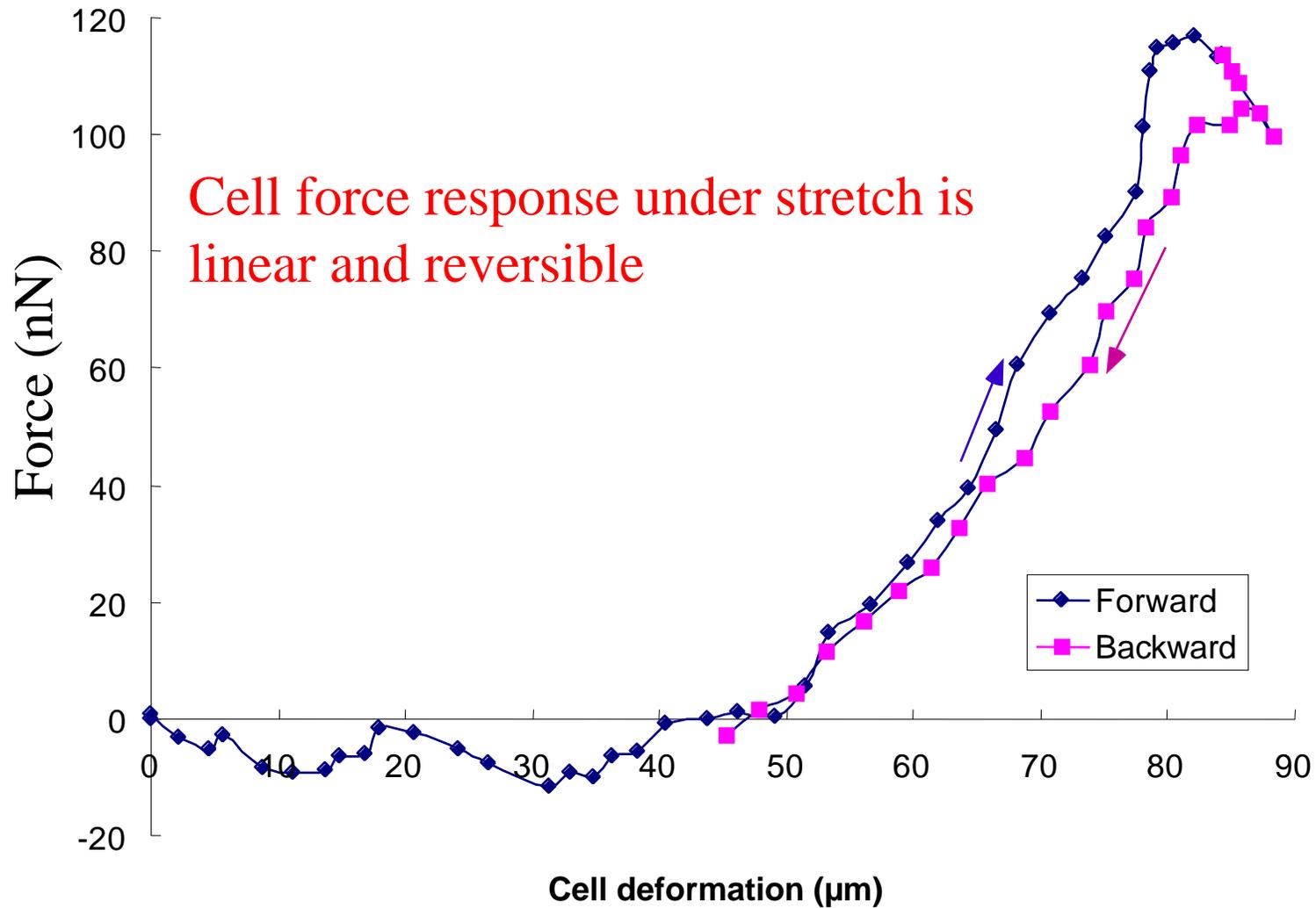
Images removed due to copyright restrictions.

## Force response of an endothelial cell



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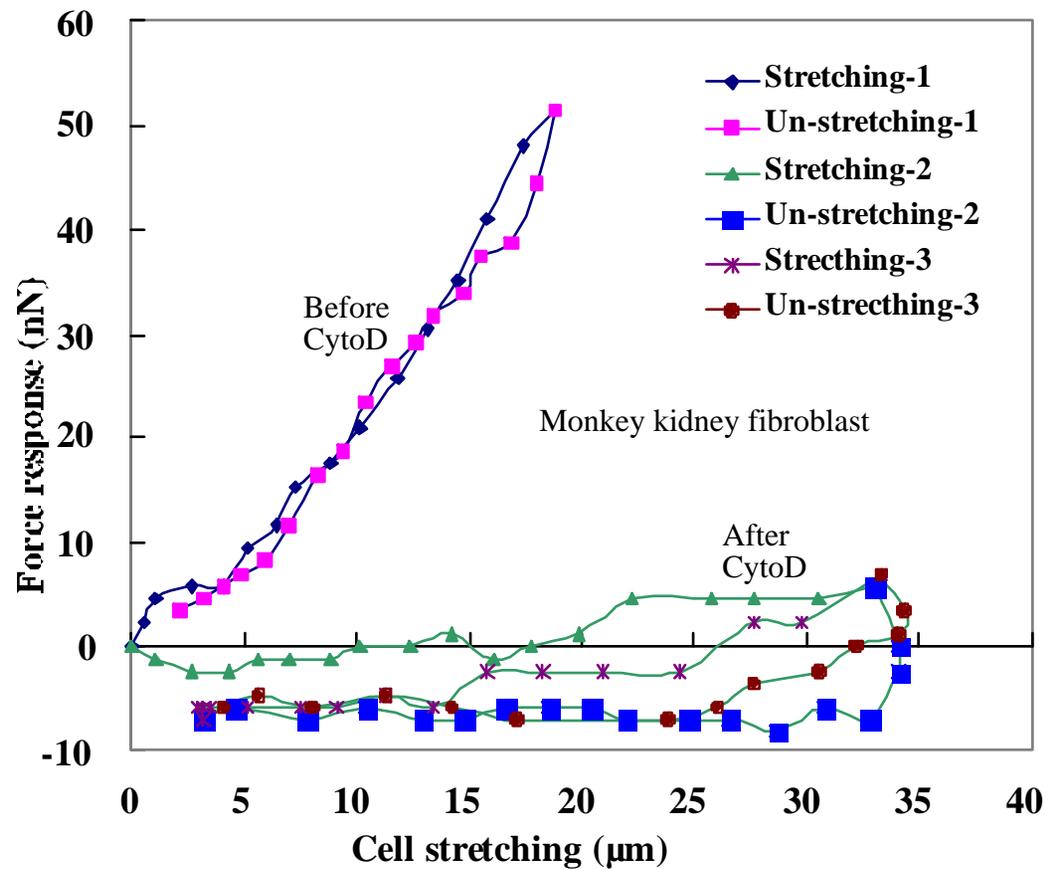
## Force response of a monkey kidney fibroblast cell



Reference: Yang and Saif. *Experimental Cell Research* 305 (2005) 42– 50.

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## Cyto-D treatment disrupts force bearing capacity



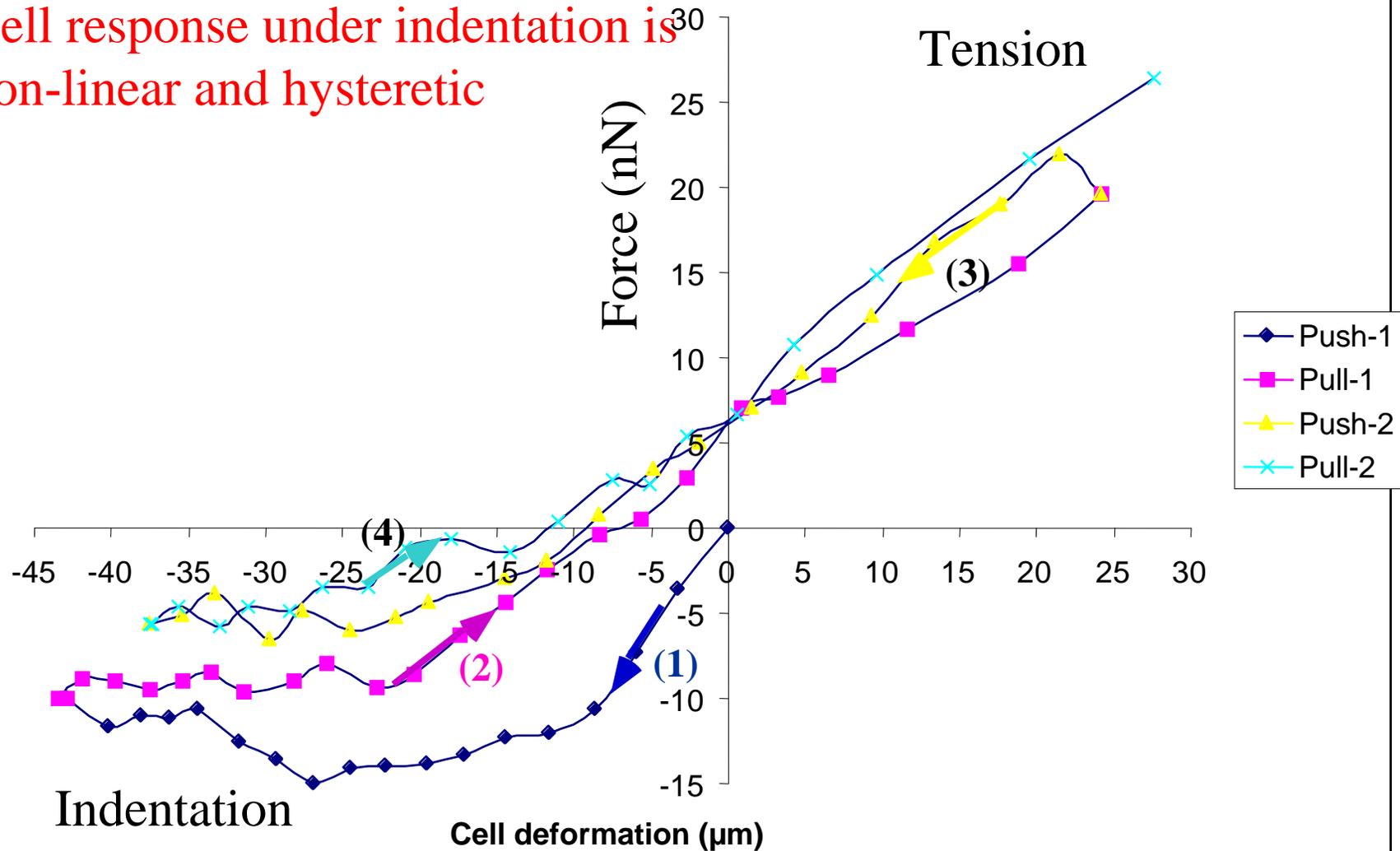
Cyto-D disrupts  
actin network

Reference: Yang and Saif. *Experimental Cell Research* 305 (2005) 42– 50.

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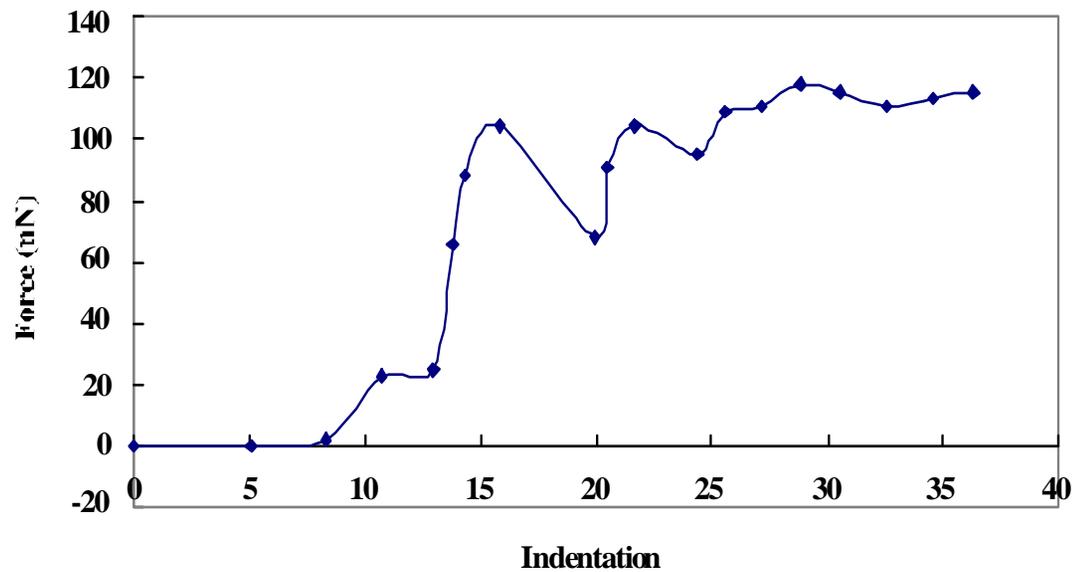
**Force response of a monkey kidney fibroblast cell**

Cell response under indentation is non-linear and hysteretic



# Mechanism of non-linearity and irreversibility under indentation

Image removed due to copyright restrictions. Photograph of GFP actin protein.



# Actin agglomerates irreversibly under indentation

Images removed due to copyright restrictions.  
Images of GFP actin undergoing indentation.

Yang and Saif  
Actabiomaterialia 2006 (in press)

## More evidence of actin agglomeration

Images removed due to copyright restrictions.

Actin in monkey kidney fibroblast is subjected to indentation.

Monkey kidney fibroblast subjected to mechanical indentation (injury simulation). Here actin stress fibers are highlighted by green fluorescent protein (GFP). In response to indentation, the cell signals local actin agglomeration at discrete locations. Such actin agglomeration is also observed in various physiological conditions such as during ischemic attack in kidney cells. This is the first evidence of actin agglomeration due to mechanical stimulus (Shengyuan and Saif, *Actabiomaterialia* 2006, in press).

# Actin agglomeration in physiological condition: ischemic attack

Image removed due to copyright restrictions.

Figure 5c in Ashworth, Sharon L., et al. "ADF/cofilin Mediates Actin Cytoskeletal Alterations in LLC-PK Cells During ATP Depletion."

*American Journal of Physiology - Renal Physiology* 284 (2003): F852-F862.

## Porcine kidney cells

Ashworth et al. *Am J. Physiol Renal Physiol* 284: F852, 2003.

## Why MEMS bio sensors:

1. Force range: 1-100 nN (natural progression from optical tweezer, magnetic beads, AFM)
1. Flexibility of design (cell contact region may be designed in a variety of fashions)
3. Large cell deformation range (sub  $\mu\text{m}$ -10s of  $\mu\text{m}$ )